



**GABRIEL MIRANDA MOREIRA**

**EFFECTS OF PREGNANCY ON QUANTITATIVE ASPECTS  
OF NUTRITION, PHYSIOLOGY AND METABOLISM OF  
BEEF HEIFERS**

**LAVRAS - MG  
2020**

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Tese apresentada à Universidade Federal de Lavras, como parte das exigências do Programa de Pós-Graduação em Zootecnia, área de concentração em Produção e Nutrição de Ruminantes, para a obtenção do título de Doutor.

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Orientador

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Coorientador

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**GABRIEL MIRANDA MOREIRA**

**EFEITOS DA PRENHEZ SOBRE ASPECTOS QUANTITATIVOS DA NUTRIÇÃO,  
FISIOLOGIA E METABOLISMO DE NOVILHAS DE CORTE**

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**LAVRAS - MG  
2020**

*À minha maior incentivadora e conselheira, minha mãe, Josiane.*

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## RESUMO

Existem poucas informações sobre o efeito da gestação e do tempo de gestação sobre o consumo, digestão total e parcial e utilização de nutrientes, redução do volume ruminal e mudanças na taxa de passagem em vacas zebuínas. No Brasil existem 55 milhões de cabeças que podem ser beneficiadas com os avanços do conhecimento na área. Portanto, esta pesquisa foi realizada com o objetivo de quantificar os efeitos da prenhez sobre a nutrição, fisiologia e metabolismo de novilhas de corte. Doze novilhas Zebu, canuladas no rúmen, foram divididas aleatoriamente em dois grupos [gestantes ( $n = 7$ ) e não gestantes ( $n = 5$ )]. Todas as novilhas receberam a mesma dieta durante o experimento. Novilhas gestantes acumularam reservas corporais (+ 35 kg) até 240 dias de gestação (DOP), quando então iniciaram a mobilização de tecidos (- 36 kg) até 286 DOP. Os consumos de matéria seca (MS), matéria orgânica (MO), proteína bruta (PB), fibra em detergente neutro (FDN) e nutrientes digestíveis totais (NDT) diminuíram ( $P < 0,06$ ) com o decorrer da gestação. A digestibilidade aparente do trato total de MS e FDN e a digestibilidade ruminal de MO e FDN foram menores ( $P \leq 0,09$ ) para novilhas gestantes em comparação com novilhas não gestantes. A digestibilidade aparente do trato total ( $P < 0,01$ ) de PB aos 267 e 286 DOP foi maior ( $P < 0,01$ ) em novilhas gestantes em comparação com não gestantes. O pool ruminal de matéria fresca (- 7,10 kg), MS (- 1,30 kg) e FDN (- 0,63 kg) foi menor ( $P < 0,02$ ) em novilhas gestantes do que em não gestantes aos 267 DOP. Em todos os períodos experimentais, a taxa de passagem da MS em novilhas gestantes foi maior ( $P < 0,09$ ) que em novilhas não gestantes. Nenhuma diferença foi encontrada no balanço de nitrogênio e nos parâmetros de fermentação ruminal. Novilhas gestantes foram mais eficientes ao longo do tempo para sintetizar proteína microbiana. A frequência cardíaca de novilhas em final de gestação em comparação com as não gestantes aumentou em oito batimentos/min quando avaliadas pouco antes da alimentação matinal, chegando a 11 batimentos/min quatro horas após a alimentação matinal. As concentrações de glicose antes da alimentação matinal foram semelhantes durante todos os períodos de coleta, com exceção daquela aos 286 dias (interação DOP  $\times$  estado fisiológico;  $P = 0,05$ ) quando a glicose foi mais baixa nas novilhas gestantes (83 mg/dL) em comparação com não gestantes (107 mg/dL). Os genes relacionados à remodelação, inflamação e transporte de ácidos graxos voláteis,  $H^+$ ,  $HCO_3^-$  e glicose no epitélio ruminal foram regulados negativamente no final da gestação. Esses resultados sugerem que o epitélio ruminal economiza energia no final da gestação para beneficiar o desenvolvimento fetal. Além disso, o aumento da frequência cardíaca e a mobilização de tecidos podem ser considerados mecanismos homeorréticos que auxiliam no atendimento das necessidades nutricionais fetais. O estado fisiológico e o estágio de gestação devem ser incluídos nos modelos de predição de desempenho, uma vez que novilhas de corte no final da gestação são menos eficientes na extração de energia da dieta em comparação com animais não gestantes, alterando os nutrientes digestíveis totais preditos na alimentação.

**Palavras-chave:** Capacidade Ruminal, Consumo de Matéria Seca, Digestibilidade Intestinal, Digestibilidade Ruminal, Epitélio Ruminal, Expressão gênica, Frequência Cardíaca, Homeorrese, Tempo de Retenção Ruminal, Zebu

## ABSTRACT

There is a lack of information about the effect of gestation and gestation time on intake, total and partial digestion and nutrient utilization, reduction of ruminal volume and changes in the feed passage rate in Zebu cows. Brazil has 55 million heads that could be benefited from the knowledge advances in this area. Therefore, this research was carried out to quantify the effects of pregnancy on the nutrition, physiology and metabolism of beef heifers. Twelve ruminally cannulated Zebu beef heifers were divided at random into two groups [pregnant (n = 7) and non-pregnant (n = 5)]. All heifers received the same diet throughout the experiment. Pregnant heifers accumulated body reserves (+ 35 kg) up to 240 days of pregnancy (DOP), then started mobilizing tissues (- 36 kg) until 286 DOP. The intake of dry matter (DM), organic matter (OM), crude protein (CP), neutral detergent fiber (NDF) and total digestible nutrients (TDN) reduces ( $P < 0.06$ ) with the course of pregnancy. The apparent total-tract digestibility of DM and NDF and ruminal digestibility of OM and NDF was lower ( $P \leq 0.09$ ) for pregnant compared with non-pregnant heifers. Crude protein apparent total-tract digestibility at 267 and 286 DOP was greater ( $P < 0.01$ ) to pregnant compared with non-pregnant heifers. The ruminal pool of wet matter (- 7.10 kg), DM (- 1.30 kg) and NDF (- 0.63 kg) was lower ( $P < 0.02$ ) to pregnant than non-pregnant heifers at 267 DOP. In all experimental periods, DM passage rate was greater ( $P < 0.09$ ) to pregnant than non-pregnant heifers. No difference was found on nitrogen balance and ruminal fermentation parameters. Pregnant heifers were more efficient over time to synthesize microbial protein. The heart rate of late-pregnant heifers compared to controls increased by eight beats/min when evaluated just before morning feeding, and the difference reached 11 beats/min when evaluated four hours after morning feeding. Glucose levels before morning feeding were similar during all collection periods with an exception at 286 days (DOP  $\times$  physiological status interaction;  $P = 0.05$ ) when glucose was lower in pregnant (83 mg/dL) compared to non-pregnant (107 mg/dL) heifers. Both genes related to remodeling, inflammation, and volatile fatty acids,  $H^+$ ,  $HCO_3^-$ , and glucose transport in the ruminal epithelium were downregulated at late gestation. These results suggest that the ruminal epithelium saves energy at late pregnancy to benefit fetal development. In addition, the increase in heart rate coupled with tissue mobilization can be considered homeorhetic mechanisms that help meet the fetal nutrient requirements. The physiological status, as well as the stage of gestation, should be included in performance prediction models since late-gestating beef heifers are less efficient at extracting energy from feed compared to non-pregnant animals, changing the feed predicted total digestible nutrients.

**Keywords:** Dry Matter Intake, Gene Expression, Heart Rate, Homeorhesis, Intestinal Digestibility, Ruminal Capacity, Ruminal Digestibility, Ruminal Epithelium, Ruminal Retention Time, Zebu Cattle

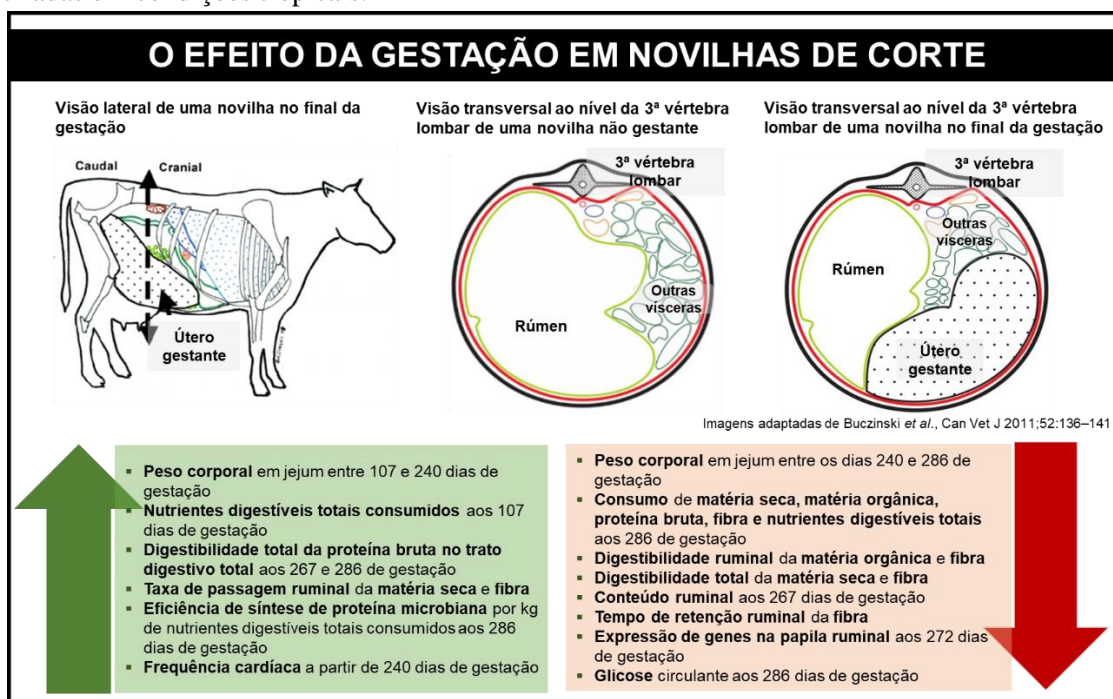


## Efeito da gestação sobre aspectos quantitativos da nutrição, fisiologia e metabolismo de novilhas de corte

Elaborado por **Gabriel Miranda Moreira** e orientado por **Mateus Pies Gionbelli**

Para formular corretamente a dieta de um animal faz-se necessário estimar sua capacidade de consumo alimentar. No entanto, o cenário observado ao final da gestação é de alta exigência nutricional e baixa capacidade de consumo. Logo, essa pesquisa foi realizada com objetivo de quantificar os efeitos da gestação sobre a nutrição, fisiologia e metabolismo de novilhas de corte.

Foram observados maior consumo de nutrientes aos 107 dias de gestação e aumento do peso corporal (+ 35 kg) até 240 dias de gestação nas novilhas gestantes. No final da gestação (> 240 dias) houve redução da capacidade ruminal e do consumo de alimentos e, conseqüentemente, perdas de peso corporal (- 36 kg) nesse período. Também foi verificado aumento da velocidade em que o alimento passa pelo rúmen (taxa de passagem) em novilhas gestantes. Entretanto, devido ao menor tempo de retenção no rúmen, tanto a digestibilidade ruminal quanto a digestibilidade total da fibra foram diminuídas. Contudo, novilhas gestantes foram mais eficientes em utilizar a energia consumida para produzir proteína microbiana. A digestão da proteína bruta no trato digestório total aumentou no final da gestação. Houve menor expressão de genes relacionados à remodelação, inflamação e transporte de ácidos graxos voláteis,  $H^+$ ,  $HCO_3^-$  e glicose no epitélio ruminal. A frequência cardíaca das novilhas gestantes foi maior no final da gestação. Portanto, concluímos que a gestação induz alterações sobre a nutrição, fisiologia e metabolismo de novilhas de corte. A quantificação dessas alterações fornece embasamento científico para a melhoria de modelos de ajuste do consumo de alimento e, conseqüentemente, melhor direcionamento das decisões a serem tomadas no sistema produtivo em relação a novilhas/vacas de corte gestantes criadas em condições tropicais.



**Efeitos da gestação em novilhas zebuínas de corte. Ao final da gestação o útero ocupa grande parte da cavidade abdominal comprimindo o rúmen induzindo alterações nutricionais, fisiológicas e metabólicas em novilhas de corte.**

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## 1 **FIRST SECTION**

### 3 **1 INTRODUCTION**

5 The Brazilian bovine herd has 213.5 million heads (INSTITUTO BRASILEIRO DE  
6 GEOGRAFIA E ESTATÍSTICA, 2019), with an estimated 80% zebu component (*Bos taurus*  
7 *indicus*), composed mainly of Nellore cattle (NASCIMENTO, 2020). Of the total herd, around  
8 55 million are breeding beef cows (FERRAZ et al., 2018). Breeding cows are permanent on the  
9 herd and have a high maintenance cost. Ritchie (1995) suggests that 50% of the total energy  
10 expended in beef production is used for maintenance. In beef cow-calf operation systems, 60 to  
11 70% of the energy demand is for lactation and reproduction. Thus, it is essential to know  
12 adequately the nutritional requirements to balance the best diet for that cow category.

13 Pregnancy is the most physiologically complex stage in the production cycle of a cow.  
14 Notably, in pregnant Zebu cows, nutritional requirements and prediction of feed intake are  
15 much less known, and the first studies are only dated from 2010 onwards (GIONBELLI, 2013).  
16 Research dealing with pregnancy in cows is more difficult because of the inherently long  
17 experimental evaluation time.

18 In order to balance a diet correctly, it is necessary to accurately predict animal feed  
19 intake. However, it is difficult to predict the ingestion capacity of pregnant cows, since physical  
20 and physiological factors are not considered in traditional ruminant feed intake regulation  
21 models. Particularities, such as the influence of the gravid uterus on the reduction of rumen  
22 capacity, hormonal regulation of pregnancy, or the homeorhetic mechanism of nutrient usage,  
23 are difficult to model and are probably the main causes of the variations in voluntary intake  
24 observed in pregnant cattle.

25 Thus, this study aimed:

- 26 a. to increase information on the effect of gestation period on the intake, digestion, and  
27 use of nutrients in Zebu beef heifers;
- 28 b. to yield accurate information on rumen volume reduction and ingesta flow rate  
29 according to the gestation length in Zebu beef heifers;
- 30 c. to generate data on the effect of gestation time on the partial digestibility of dry matter  
31 and other dietary components in the rumen and intestine in Zebu beef heifers.
- 32 d. to investigate possible metabolic and physiological changes induced by pregnancy,  
33 as well as the behavior of important genes linked to the ruminal epithelium activity  
34 towards the advance of gestation in Zebu beef heifers.

35 The information obtained in this experiment may help to provide scientific basis to  
36 propose improved models for adjusting feed intake and, consequently, improving decision  
37 making in production systems relating to pregnant beef heifers/cows under tropical conditions.  
38

## 39 **2 BACKGROUND**

40  
41 During cow pregnancy, several challenges occur to the normal physiological status to  
42 support and allocate resources for the conception and development of a new individual  
43 (GIONBELLI *et al.*, 2015). Beef cows have a dramatic increase in nutrient demand during late  
44 gestation. To illustrate this, the maintenance requirements for metabolizable energy (ME)  
45 increase 28% in the last 90 days of pregnancy (DOP), considering a 500 kg zebu beef cow with  
46 no variation in body condition score (VALADARES FILHO *et al.*, 2016). Concurrently with  
47 nutritional demand increases, feed intake seems to decrease, leading to the establishment of  
48 additional mechanisms to maintain pregnancy and development of a healthy calf.

49 Fundamentally, pregnancy in cows impacts metabolism through changes in feed intake,  
50 ruminal fill, digesta passage rate, feed digestibility, ruminal fermentation parameters, maternal  
51 body weight (BW), circulating serum metabolites, and steroid hormones.  
52

### 53 **2.1 Effects of pregnancy on feed intake and ruminal capacity**

54  
55 The advancement of gestation may reduce feed intake in ruminant females and, during  
56 the days closer to parturition, the reduction in feed intake is even more drastic (FORBES, 2007).  
57 Initially, the reduction in feed intake observed in the late gestation of ruminants was related to  
58 ruminal compression generated by the growth of the gravid uterus (MAKELA, 1956). This  
59 association became known as Makela's compression theory. In the 1960s, several studies were  
60 carried out to assess intake and possible factors that can change it during pregnancy (BROSTER  
61 *et al.*, 1964; CAMPLING, 1966; FORBES, 1968; GRAHAM; WILLIAMS, 1962; JOHNSON  
62 *et al.*, 1966; LAMBERTH, 1969; REID; HINKS, 1962). At the end of the decade, the  
63 conclusion was that pregnancy had a physical impact on ruminal capacity. However, intake was  
64 not always changed or the variation was insignificant, where changes in the passage rate of  
65 digesta, as well as metabolic and hormonal mechanisms, were suggested as possible regulators  
66 of feed intake.

67 Until the end of the 20th century, more studies showed the effect of pregnancy on the  
68 intake and nutrients use in ruminants (COFFEY *et al.*, 1989; FORBES, 1970; FORBES, 1986;

69 GUNTER *et al.*, 1990; HANKS *et al.*, 1993; VANZANT *et al.*, 1991). Feed intake is directly  
70 related to rumen capacity (MERTENS, 1987), although during pregnancy this effect is not  
71 entirely clear. Hanks *et al.* (1993) verified lower gastrointestinal fill in pregnant than in non-  
72 pregnant limit-fed beef cows during the last trimester of pregnancy. Similarly, Stanley *et al.*  
73 (1993) using pregnant beef cows given *ad libitum* access to feed, reported a decrease in ruminal  
74 fill between 65 and 24 days prepartum, whereas dry matter (DM) intake increased. This  
75 information provides sufficient evidence to indicate that there is a physical effect of gestation  
76 on the reduction of ruminal capacity of ruminants at late gestation. Forbes (1986) remember  
77 that the effects of physical compression coincide with the changes in endocrine factors, such as  
78 estrogen levels, and body reserves, mediated in response to the advancement of gestation and  
79 preparation for the future lactation.

80 In the last 20 years, researchers remain evaluating the effect of pregnancy on feed intake  
81 (HARE *et al.*, 2019; LINDEN *et al.*, 2014; ROTTA *et al.*, 2015b; SCHEAFFER *et al.*, 2001;  
82 WOOD *et al.*, 2013). Scheaffer *et al.* (2001) showed that pregnant crossbred beef heifers had  
83 lower total ruminal fill (g/kg BW) than non-pregnant heifers at 200 and 270 DOP. However,  
84 DM intake and ME intake did not change from early-to-late gestation. Similarly, Wood *et al.*  
85 (2013) did not found difference in DM intake between pregnant and non-pregnant mature beef  
86 cows (Angus and Simmental cross-breeding). Conversely, Holstein × Gyr dairy cows decreased  
87 DM intake from 50 days before parturition (ROTTA *et al.*, 2015b). Also, Hare *et al.* (2019)  
88 verified that pregnant Hereford cross heifers increased DM intake between eight to two weeks  
89 prepartum by 18%, then decreased it by 8.0% in the week before parturition.

90 The effects of pregnancy on DM intake of forage-fed beef heifers were compared to  
91 mature beef cows (LINDEN *et al.*, 2014), where over the seven weeks prepartum, DM intake  
92 as a percentage of BW was similar between cows and heifers. Moreover, a tendency for an age  
93 × pregnancy × time interaction was also observed for DM intake once pregnant heifers  
94 demonstrated increases in intake from seven through two weeks prepartum followed by a small  
95 decline in the final week. However, pregnant cows demonstrated relatively stable intakes for  
96 most of the prepartum period with a minor increase in the final week.

97 Therefore, the effect of pregnancy on voluntary feed intake of ruminant depends on an  
98 interaction between physical and physiological factors, rendering intake estimation a complex  
99 task. The third edition of Nutrient Requirements of Zebu and Crossbred cattle (BR-CORTE  
100 3.0) proposes a reduction of 0.0204 grams of DM per kg shrunk body weight (SBW) for each  
101 day over 135 DOP by pregnant zebu cows (VALADARES FILHO *et al.*, 2016). Another key  
102 point is that most of the studies investigating the effect of pregnancy on feed intake were carried

103 out in temperate regions and with taurine animals, and information for zebu cattle raised in  
104 tropical regions is still lacking.

105

## 106 **2.2 Effects of pregnancy on passage rate of digesta and feed digestibility**

107

108 The main justification for absence of a reduction in feed intake during pregnancy is the  
109 increase in the digesta passage rate. Passage rate is a measure of the time by which a portion of  
110 digesta is exposed to the processes of mixing, digestion, and absorption in the gastrointestinal  
111 tract or a defined segment; it is measured as the mean retention time, which is the ratio of the  
112 amount of any component of digesta in a segment to the flow of that digesta component from  
113 that segment (DIJKSTRA *et al.*, 2005). Passage rates are affected by a wide variety of factors  
114 that have different effects, including animal and feed factors (MOYO; NSAHLAI, 2018).

115 Most of the published studies propose that movement of digesta through the  
116 gastrointestinal tract of pregnant ruminants is increased such that it compensates for the loss of  
117 ruminal capacity caused by fetal growth in late gestation (ewes: (COFFEY *et al.*, 1989;  
118 FAICHNEY; WHITE, 1988; KASKE; GROTH, 1997; WESTON, 1988); beef cows: (HANKS  
119 *et al.*, 1993; LINDEN *et al.*, 2014; STANLEY *et al.*, 1993; VANZANT *et al.*, 1991). Okine  
120 and Mathison (1991) indicated that the ruminal passage rate of neutral detergent fiber (NDF)  
121 increased concomitantly with intake. But, during late gestation feed intake seems to be not the  
122 only factor that changes passage rate. Pregnant cows and heifers fed *ad libitum* or on the pasture  
123 had faster digesta passage rates than their non-pregnant counterparts (LINDEN *et al.*, 2014;  
124 STANLEY *et al.*, 1993; VANZANT *et al.*, 1991). Likewise, pregnant ewes fed *ad libitum*  
125 decreased mean retention time of digesta compared to non-pregnant ewes (COFFEY *et al.*,  
126 1989; KASKE; GROTH, 1997). On the other hand, Hanks *et al.* (1993) found that limit-fed  
127 pregnant cows had increased particulate passage rate and decreased ruminal and total-tract  
128 mean retention time when compared to non-pregnant cows. Likewise, an increased passage rate  
129 of digesta from the reticulorumen has also been demonstrated for late pregnancy ewes fed at a  
130 constant level throughout pregnancy (FAICHNEY; WHITE, 1988; GUNTER *et al.*, 1990;  
131 WESTON, 1988). With this in mind, it can be concluded that the impact of gestation on the  
132 passage rate of digesta occurs through factors other than feed intake. Neural or hormonal factors  
133 might contribute to an increase in prepartum passage rates (STANLEY *et al.*, 1993). According  
134 to Forbes (1986), the circulating estrogen levels may increase passage rate.

135 Feed digestibility and passage rate of digesta are inversely proportional, in other words,  
136 decreases in apparent total tract digestibility of diet components may be explained in part by

137 the increase in passage rate (COLUCCI *et al.*, 1982; RIBEIRO *et al.*, 2015). Although the  
138 passage rate increases during gestation, studies evaluating the effects of pregnancy on  
139 digestibility in cattle are not consistent. Scheaffer *et al.* (2001) reported lower *in vitro* DM  
140 digestibility in pregnant heifers when compared with non-pregnant heifers. Vanzant *et al.*  
141 (1991) and Hanks *et al.* (1993) found no difference in organic matter (OM) and DM  
142 digestibility, respectively, between pregnant and non-pregnant animals. On the other hand,  
143 Linden *et al.* (2014) noted that DM digestibility was greater for pregnant animals than for non-  
144 pregnant animals and decreased more overtime for non-pregnant than for pregnant animals.  
145 However, these results represent apparent total-tract digestibility which is the sum of ruminal,  
146 and post-ruminal digestibilities. Therefore, it is necessary to establish more clearly the  
147 relationship between feed digestibility and passage rate in pregnant cows, mainly zebu cattle,  
148 as well as to assess changes in partial digestions (ruminal and intestinal) as a function of  
149 pregnancy time.

150

### 151 **2.3 Effects of pregnancy on ruminal parameters and microbial protein synthesis**

152

153 Since pregnancy can cause changes in feed intake, passage rate, and digestibility, it is  
154 expected that the ruminal parameters should also change. For instance, ruminal pH depends on  
155 saliva production, volatile fatty acids (VFA) balance, type and level of feed intake, and on the  
156 exchange of bicarbonates and phosphates through the ruminal epithelium (ASCHENBACH *et al.*  
157 *et al.*, 2011). Rumen VFA concentration reflects the balance between production and clearance  
158 by passage with the fluid phase into the omasum or by absorption through the ruminal wall  
159 (LOPEZ *et al.*, 2003). Additionally, rumen ammoniacal nitrogen (N-NH<sub>3</sub>) is a very potent  
160 buffer once NH<sub>3</sub> can immediately bind H<sup>+</sup> to form NH<sub>4</sub><sup>+</sup> in the ruminal content. Ruminal N-  
161 NH<sub>3</sub> removal occurs through the use of ruminal bacteria, efflux to the omasum, or absorption  
162 across the ruminal wall (ASCHENBACH *et al.* 2011).

163 Hanks *et al.* (1993) found that ruminal N-NH<sub>3</sub> did not differ between pregnant and non-  
164 pregnant cows until 10 days before parturition when pregnant cows (6.9 mg/dL) had lower  
165 concentrations than non-pregnant cows (8.0 mg/dL). The same authors also observed that total  
166 VFA concentration was lower in pregnant than non-pregnant cows at 203 DOP; however, this  
167 relationship was reversed at 230 DOP. On the other hand, ruminal pH and VFA concentration  
168 did not differ between pregnant and non-pregnant heifers, while ruminal N-NH<sub>3</sub> was reduced  
169 due to pregnancy (SCHEAFFER *et al.*, 2001). Besides, Hare *et al.* (2019) verified that ruminal  
170 N-NH<sub>3</sub> had a treatment × day interaction, where pregnant animals fed excess (+ 33%)

171 metabolizable protein (MP) decreased as parturition approached (10.1 to 8.6 mg/ dL); whereas,  
172 N-NH<sub>3</sub> was not affected for pregnant animals fed to meet MP requirements (1.0 to 1.3 mg/dL).

173 Nitrogen (N) availability and its synchronization with energy sources are the greatest  
174 determinant of the amount of microbial crude protein (MCP) synthesized in the rumen (CLARK  
175 *et al.*, 1992). Conceptually, microbial efficiency is the amount of MCP obtained from a  
176 determined energy unit, or else, it is the amount of protein produced by ruminal microorganisms  
177 from energy substrates available in the rumen, under the interference of a series of factors  
178 (SANTOS *et al.*, 2016). However, there is a lack of studies evaluating the direct effects of  
179 pregnancy on MCP synthesis in the literature. In conditions where the passage rate is high, there  
180 is an expected reduction in microbial maintenance costs due to a reduction in the ruminal  
181 retention time (GIONBELLI, 2013).

182

#### 183 **2.4 Effects of pregnancy on maternal body weight**

184

185 According to Bauman and Currie (1980), metabolism control during pregnancy besides  
186 homeostasis also involves homeorhesis regulations. These authors define homeorrhesis as  
187 orchestrated control in the metabolism of body tissues necessary to support a physiological  
188 state. For example, nutrient partitioning during pregnancy prioritizes fetal growth (fetus and  
189 fetal membranes) and gravid uterus as well as the development of the mammary gland  
190 (BAUMAN; CURRIE, 1980).

191 During pregnancy, and especially in its last trimester, nutrient requirements increase  
192 significantly, while rumen capacity is reduced due to fetal growth. Lower ruminal volume leads  
193 to lower feed intake capacity. To support the high nutritional demand, pregnant cows increase  
194 digesta passage rate, and consequently intake increases. However, this mechanism does not  
195 always meet the requirements, and tissue mobilization must take place in order to fulfill  
196 pregnancy demands. Nevertheless, Vanzant *et al.* (1991) observed that pregnant heifers were  
197 50 kg heavier than non-pregnant at 55 and 12 days before calving. However, BW assessment  
198 may not accurately represent maternal tissues weights due to the growth of pregnancy-related  
199 components, such as the gravid uterus and mammary gland (GIONBELLI *et al.*, 2015).  
200 Scheaffer *et al.* (2001) investigated the impact of pregnancy and advancing gestation on BW in  
201 crossbred beef heifers. The carcass weight and eviscerated BW of pregnant heifers were greater  
202 at 200 DOP, but the carcass weight of pregnant compared with that of non-pregnant heifers  
203 tended to be lower at 270 DOP. In another study, cows and heifers were fed tallgrass prairie  
204 hay (low nutritional quality) for *ad libitum* intake and 450 g/day of soybean meal. It was



205 observed that over the prepartum phase (7 to 1 week before calving) BW decreased more in  
206 pregnant than in non-pregnant cows, indicating that the low-quality forage did not meet the  
207 energy requirements of pregnancy even with protein supplementation from soybean meal  
208 (LINDEN *et al.*, 2014). Hare *et al.* (2019) observed that increasing MP supply by 33% over  
209 predicted requirements improved N balance and decrease indicators of skeletal muscle  
210 catabolism, hence increasing BW gain during late gestation. Some authors suggested that to  
211 compensate restricted diets pregnant ruminants may alter the maternal visceral organ mass to  
212 benefit the offspring (MEYER *et al.*, 2010; REED *et al.*, 2007; ROTTA *et al.*, 2015a;  
213 SCHEAFFER *et al.*, 2001; WOOD *et al.*, 2013).

214

## 215 **2.5 Effects of pregnancy on circulating serum metabolites and steroid hormones**

216

217 Circulating serum metabolites are used as indicators of the nutritional and metabolic  
218 status of the dam. Beef cows may alter metabolism in response to the increase in nutrient  
219 demands especially during mid- to late gestation (WOOD *et al.*, 2013). Glucose is a major  
220 energy source used for both maternal tissues as well as fetal growth and development during  
221 pregnancy (DOORNENBAL; TONG; MURRAY, 1984). During late gestation, there is an  
222 increase in hepatic gluconeogenesis along with decreased glucose utilization by tissues (BELL,  
223 1995). In beef cows and heifers, plasma glucose concentrations were lower in pregnant than in  
224 non-pregnant animals in the last 50 DOP, likely due to glucose use by the fetus according to  
225 Linden *et al.* (2014). In contrast, glucose concentrations increase during the last week of  
226 pregnancy in dairy cows (INGVARTSEN; ANDERSEN, 2000).

227 Circulating non-esterified fatty acid (NEFA) and beta-hydroxybutyrate (BHB)  
228 concentrations are indicators of fat mobilization and ketogenic metabolic status. Bell (1995)  
229 suggests that increased BHB as a result of incomplete NEFA oxidation. In mid-to-late gestation,  
230 pregnant cows had increased circulating NEFA, and BHB concentrations and reduced  
231 circulating total cholesterol concentrations when compared to non-pregnant cows, indicative of  
232 greater fat catabolism and a more ketogenic metabolic status during pregnancy (WOOD *et al.*,  
233 2013). Similarly, pregnant cows and heifers had higher plasma BHB than their non-pregnant  
234 counterparts (LINDEN *et al.*, 2014). This observation is justified by insufficient dietary energy,  
235 as a result of low feed quality, during pregnancy leading to increased lipolysis.

236 Prepartum changes in feed intake and digesta passage rate are more consistent with  
237 changes in blood concentrations of steroid hormones. Grummer *et al.* (1990) speculated that  
238 the surge in blood estrogen might be responsible for the depression in feed intake before

239 parturition. Plasma estrogen concentrations increase to around tenfold to twentyfold at about  
240 one month before calving (FORBES, 2007). Studies in rats also indicate that the effect of  
241 estrogen on feed intake is due, at least in part, to direct actions in the brain, where the  
242 paraventricular nucleus of the hypothalamus is the main site of action for estrogen on feed  
243 intake (BUTERA; BEIKIRCH, 1989). Estrogen has been proposed to interact with orexigenic  
244 (neuropeptide Y, ghrelin, and melanin-concentrating hormone) and anorexigenic (insulin,  
245 leptin, serotonin, and cholecystokinin) neuropeptides influencing feed intake (BROWN;  
246 CLEGG, 2010). For example, neuropeptide Y (NPY) is a potent orexigenic, which increases  
247 feeding behavior in fed and fasted animals. Estrogen acts via the estrogen receptors in the  
248 hypothalamus to reduce feed intake and may mediate its anorectic effects by decreasing NPY  
249 expression or release. Conversely, injection of estradiol 17 $\beta$  at day 276 of gestation to achieve  
250 blood concentrations similar to those at parturition did not depress feed intake (BREMNER et  
251 al., 1999). This indicated that other hormones besides estrogen must be involved in regulating  
252 prepartum intake because the decrease in feed intake is initiated before the rise in estrogen  
253 before parturition. Progesterone does not seem to have a direct effect on feed intake in cattle,  
254 but, since it blocks estrogen effects it may reduce its effects on feed intake.

255         The faster passage rate also is linked to circulating estrogen. Forbes (1986) reviewed  
256 the literature regarding the effects of sex hormones on voluntary intake and concluded that  
257 decreased ruminal retention time of particles during the last trimester of pregnancy probably  
258 resulted from high circulating estrogen concentrations. Furthermore, both estrogen and  
259 progesterone increase gut motility in non-pregnant cattle and sheep (FORBES, 1986). Indeed,  
260 these hormones exhibited greater concentrations in pregnant than in non-pregnant cows,  
261 coinciding with greater particulate passage rate and lower gastrointestinal mean retention time,  
262 ruminal retention time, and intestinal transit time to pregnant animals (HANKS et al., 1993).

263

## REFERENCES

- 264  
265  
266 ASCHENBACH, J. R. *et al.* Ruminant Nutrition Symposium: Role of fermentation acid  
267 absorption in the regulation of ruminal pH. **Journal of Animal Science**, 89, n. 4, p. 1092-1107,  
268 Apr. 2011.
- 269  
270 BAUMAN, D. E.; CURRIE, W. B. Partitioning of nutrients during pregnancy and lactation: a  
271 review of mechanisms involving homeostasis and homeorhesis. **Journal of Dairy Science**, 63,  
272 n. 9, p. 1514-1529, 1980.
- 273  
274 BROSTER, W. *et al.* Experiments on the nutrition of the dairy heifer: V. Nutrition in late  
275 pregnancy. **The Journal of Agricultural Science**, 63, n. 1, p. 51-58, 1964.
- 276  
277 CAMPLING, R. C. A preliminary study of the effect of pregnancy and of lactation on the  
278 voluntary intake of food by cows. **British Journal of Nutrition**, 20, n. 1, p. 25-39, 1966.
- 279  
280 CLARK, J. *et al.* Microbial protein synthesis and flows of nitrogen fractions to the duodenum  
281 of dairy cows. **Journal of Dairy Science**, 75, n. 8, p. 2304-2323, 1992.
- 282  
283 COFFEY, K. *et al.* The influence of pregnancy and source of supplemental protein on intake,  
284 digestive kinetics and amino acid absorption by ewes. **Journal of Animal Science**, 67, n. 7, p.  
285 1805-1814, 1989.
- 286  
287 COLUCCI, P. *et al.* Feed intake, apparent diet digestibility, and rate of particulate passage in  
288 dairy cattle. **Journal of Dairy Science**, 65, n. 8, p. 1445-1456, 1982.
- 289  
290 DIJKSTRA, J. *et al.* **Quantitative aspects of ruminant digestion and metabolism**. Cabi,  
291 2005. 1845931459.
- 292  
293 FAICHNEY, G.; WHITE, G. Rates of passage of solutes, microbes and particulate matter  
294 through the gastro-intestinal tract of ewes fed at a constant rate throughout gestation.  
295 **Australian Journal of Agricultural Research**, 39, n. 3, p. 481-492, 1988.
- 296  
297 FERRAZ, J. *et al.* Impact of using artificial insemination on the multiplication of high genetic  
298 merit beef cattle in Brazil. **Animal Reproduction**, 9, n. 3, p. 133-138, 2018.
- 299  
300 FORBES, J. M. The physical relationships of the abdominal organs in the pregnant ewe. **The**  
301 **Journal of Agricultural Science**, 70, n. 2, p. 171-177, 1968.
- 302  
303 FORBES, J. M. Voluntary food intake of pregnant ewes. **Journal of Animal Science**, 31, n. 6,  
304 p. 1222-1227, 1970.
- 305  
306 FORBES, J. M. The effects of sex hormones, pregnancy and lactation on digestion, metabolism  
307 and voluntary food intake. **Control of digestion and metabolism in ruminants**, 1986.
- 308  
309 FORBES, J. M. **Voluntary food intake and diet selection in farm animals**. Cabi, 2007.  
310 184593279X.
- 311  
312 GIONBELLI, M. P. **Nutrient requirements and quantitative aspects of growth,**  
313 **development and digestion of pregnant and non-pregnant Nellore cows**. 2013. Tese

- 314 (Doutorado em Zootecnia) – Universidade Federal de Viçosa, Viçosa, 2013. Disponível em:  
315 [https://www.locus.ufv.br/bitstream/handle/123456789/1870/texto%20completo.pdf?sequence](https://www.locus.ufv.br/bitstream/handle/123456789/1870/texto%20completo.pdf?sequence=1&isAllowed=y)  
316 [=1&isAllowed=y](https://www.locus.ufv.br/bitstream/handle/123456789/1870/texto%20completo.pdf?sequence=1&isAllowed=y). Acesso em: 18 jan 2020.
- 317  
318 GIONBELLI, M. P. *et al.* Achieving body weight adjustments for feeding status and pregnant  
319 or non-pregnant condition in beef cows. **PLoS One**, 10, n. 3, p. e0112111, 2015.
- 320  
321 GRAHAM, N. M.; WILLIAMS, A. The effects of pregnancy of the passage of food through  
322 the digestive tract of sheep. **Australian Journal of Agricultural Research**, Australian Journal  
323 of Agricultural Research, 13, n. 5, p. 894-900, 1962.
- 324  
325 GUNTER, S. *et al.* Digesta kinetics, ruminal fermentation characteristics and serum metabolites  
326 of pregnant and lactating ewes fed chopped alfalfa hay. **Journal of Animal Science**, 68, n. 11,  
327 p. 3821-3831, 1990.
- 328  
329 HANKS, D. R. *et al.* Effects of pregnancy on digesta kinetics and ruminal fermentation in beef  
330 cows. **Journal of Animal Science**, 71, n. 10, p. 2809-2814, Oct. 1993.
- 331  
332 HARE, K. S. *et al.* Oversupplying metabolizable protein in late gestation for beef cattle: effects  
333 on prepartum BW, ruminal fermentation, nitrogen balance, and skeletal muscle catabolism.  
334 **Journal of Animal Science**, 97, n. 1, p. 407-423, Jan. 1 2019.
- 335  
336 INSTITUTO BRASILEIRO DE GEOGRAFIA E ESTATÍSTICA. **Pesquisa da Pecuária**  
337 **Municipal 2018**, Rio de Janeiro, 26 set. 2018. Disponível em:  
338 [https://biblioteca.ibge.gov.br/visualizacao/periodicos/84/ppm\\_2018\\_v46\\_br\\_informativo.pdf](https://biblioteca.ibge.gov.br/visualizacao/periodicos/84/ppm_2018_v46_br_informativo.pdf).  
339 Acesso em: 14 fev. 2020.
- 340  
341 JOHNSON, W. *et al.* Voluntary intake of forage by Holstein cows as influenced by lactation,  
342 gestation, body weight, and frequency of feeding. **Journal of Dairy Science**, 49, n. 7, p. 856-  
343 864, 1966.
- 344  
345 KASKE, M.; GROTH, A. Changes in factors affecting the rate of digesta passage during  
346 pregnancy and lactation in sheep fed on hay. **Reproduction Nutrition Development**, 37, n. 5,  
347 p. 573-588, 1997.
- 348  
349 LAMBERTH, J. The effect of pregnancy in heifers on voluntary intake, total rumen contents,  
350 digestibility and rate of passage. **Australian Journal of Experimental Agriculture**, 9, n. 40,  
351 p. 493-496, 1969.
- 352  
353 LINDEN, D. *et al.* Effects of gestation and lactation on forage intake, digestion, and passage  
354 rates of primiparous beef heifers and multiparous beef cows. **Journal of Animal Science**, 92,  
355 n. 5, p. 2141-2151, 2014.
- 356  
357 LOPEZ, S. *et al.* Effects of volatile fatty acid supply on their absorption and on water kinetics  
358 in the rumen of sheep sustained by intragastric infusions. **Journal of Animal Science**, 81, n.  
359 10, p. 2609-2616, 2003.
- 360  
361 MAKELA, A. Studies on the question of bulk in the nutrition of farm animals with special  
362 reference to cattle. **Acta Agralia Fennica**, 1956.
- 363

- 364 MERTENS, D. Predicting intake and digestibility using mathematical models of ruminal  
365 function. **Journal of Animal Science**, 64, n. 5, p. 1548-1558, 1987.  
366
- 367 MEYER, A. M. *et al.* Effects of stage of gestation and nutrient restriction during early to mid-  
368 gestation on maternal and fetal visceral organ mass and indices of jejunal growth and  
369 vascularity in beef cows. **Journal of Animal Science**, 88, n. 7, p. 2410-2424, July 2010.  
370
- 371 MOYO, M.; NSAHLAI, I. V. Rate of Passage of Digesta in Ruminants; Are Goats Different?  
372 *In: Goat Science*, 2018. cap. Chapter 3.  
373
- 374 NASCIMENTO, S. **Como o zebu ajudou o Brasil a formar um rebanho de alta genética.**  
375 *Revista Globo Rural*, Uberaba, 22 abr. 2019. Disponível em:  
376 [https://revistagloborural.globo.com/Noticias/Criacao/Boi/noticia/2019/04/como-o-zebu-](https://revistagloborural.globo.com/Noticias/Criacao/Boi/noticia/2019/04/como-o-zebu-ajudou-o-brasil-formar-um-rebanho-de-alta-geneticasite.html)  
377 [ajudou-o-brasil-formar-um-rebanho-de-alta-geneticasite.html](https://revistagloborural.globo.com/Noticias/Criacao/Boi/noticia/2019/04/como-o-zebu-ajudou-o-brasil-formar-um-rebanho-de-alta-geneticasite.html). Acesso em: 14 fev. 2020.  
378
- 379 OKINE, E. K.; MATHISON, G. W. Reticular contraction attributes and passage of digesta from  
380 the ruminoreticulum in cattle fed roughage diets. **Journal of Animal Science**, 69, n. 5, p. 2177-  
381 2186, May 1991.  
382
- 383 REED, J. *et al.* Effects of selenium supply and dietary restriction on maternal and fetal body  
384 weight, visceral organ mass and cellularity estimates, and jejunal vascularity in pregnant ewe  
385 lambs. **Journal of Animal Science**, 85, n. 10, p. 2721-2733, 2007.  
386
- 387 REID, R.; HINKS, N. Studies on the carbohydrate metabolism of sheep. XVII. Feed  
388 requirements and voluntary feed intake in late pregnancy, with particular reference to  
389 prevention of hypoglycaemia and hyperketonaemia. **Australian Journal of Agricultural**  
390 **Research**, 13, n. 6, p. 1092-1111, 1962.  
391
- 392 RIBEIRO, R. *et al.* Effects of roughage sources produced in a tropical environment on forage  
393 intake, and ruminal and microbial parameters. **Journal of Animal Science**, 93, n. 5, p. 2363-  
394 2374, 2015.  
395
- 396 RITCHIE, H. D., 1995, **The optimum cow: what criteria must she meet?** Beef Improvement  
397 Federation Sheridan.  
398
- 399 ROTTA, P. *et al.* Effects of day of gestation and feeding regimen in Holstein× Gyr cows: II.  
400 Maternal and fetal visceral organ mass. **Journal of Dairy Science**, 98, n. 5, p. 3211-3223,  
401 2015a.  
402
- 403 ROTTA, P. *et al.* Effects of day of gestation and feeding regimen in Holstein x Gyr cows: I.  
404 Apparent total-tract digestibility, nitrogen balance, and fat deposition. **Journal of Dairy**  
405 **Science**, 98, n. 5, p. 3197-3210, May 2015b.  
406
- 407 SANTOS, S. A. *et al.* Protein ruminal degradation of feeds and microbial protein synthesis. In:  
408 VALADARES FILHO, S. de C. *et al.* **Nutrient Requirements of Zebu and Crossbred Cattle**  
409 **- BR-CORTE**. Viçosa: Suprema, 2016. p. 43-84  
410
- 411 SCHEAFFER, A. *et al.* Influence of pregnancy on body weight, ruminal characteristics, and  
412 visceral organ mass in beef heifers. **Journal of Animal Science**, 79, n. 9, p. 2481-2490, 2001.  
413

- 414 STANLEY, T. *et al.* Periparturient changes in intake, ruminal capacity, and digestive  
415 characteristics in beef cows consuming alfalfa hay. **Journal of Animal Science**, 71, n. 3, p.  
416 788-795, 1993.
- 417
- 418 VALADARES FILHO, S. de C. *et al.* **Nutrient Requirements of Zebu and Crossbred Cattle**  
419 **- BR-CORTE**. 3. ed. Viçosa: Suprema, 2016. 314 p.
- 420
- 421 VANZANT, E. S. *et al.* Pregnancy and lactation in beef heifers grazing tallgrass prairie in the  
422 winter: influence on intake, forage utilization, and grazing behavior. **Journal of Animal**  
423 **Science**, 69, n. 7, p. 3027-3038, Jul 1991.
- 424
- 425 WESTON, R. Factors limiting the intake of feed by sheep. 11. The effect of pregnancy and  
426 early lactation on the digestion of a medium-quality roughage. **Australian Journal of**  
427 **Agricultural Research**, 39, n. 4, p. 659-669, 1988.
- 428
- 429 WOOD, K. *et al.* Influence of pregnancy in mid-to-late gestation on circulating metabolites,  
430 visceral organ mass, and abundance of proteins relating to energy metabolism in mature beef  
431 cows. **Journal of Animal Science**, 91, n. 12, p. 5775-5784, 2013.

432 **SECOND SECTION – ARTICLE**

433

434 **ARTICLE 1 - Pregnancy affects maternal performance, ruminal digestibility,**  
435 **digesta passage rate, and efficiency of microbial protein synthesis in zebu beef**  
436 **heifers**

437

438 Article formatted according to Livestock Science guidelines

439

440 **Pregnancy affects maternal performance, ruminal digestibility, digesta passage**  
441 **rate, and efficiency of microbial protein synthesis in zebu beef heifers**

442

443 **Abstract**

444 The aim was to quantify the effects of physiological status (PS; pregnant and non-  
445 pregnant) and pregnancy time on maternal performance, feed intake, digestibility and  
446 digestion kinetics in zebu beef heifers. Twelve rumen-cannulated Zebu beef heifers (7  
447 pregnant and 5 non-pregnant) receiving the same diet during all pregnancy until  
448 calving were assigned to an experimental design. Samples were obtained at six  
449 collection periods throughout gestation [107, 170, 208, 240, 267, and 286 days of  
450 pregnancy (DOP)]. Pregnant heifers accumulated body reserves (+ 35 kg) from 107  
451 until 240 DOP, but diminished body weight (- 36 kg) from this point until 286 DOP. Dry  
452 matter (DM), organic matter (OM), crude protein (CP), neutral detergent fiber (NDF)  
453 and total digestible nutrients (TDN) intake decreased ( $P < 0.06$ ) as pregnancy  
454 progressed. Apparent total-tract digestibilities of DM and NDF and ruminal  
455 digestibilities of OM and NDF were lower ( $P \leq 0.09$ ) in pregnant compared with non-  
456 pregnant heifers. There was an interaction effect ( $P = 0.04$ ) between physiological

457 state (PS) and DOP on CP apparent total-tract digestibility. Crude protein digestibility  
458 was greater ( $P < 0.01$ ) at 267 and 286 DOP in pregnant compared to non-pregnant  
459 heifers. Intestinal digestibility of diet components was similar independently of PS.  
460 Ruminant pool of wet matter (- 7.10 kg), DM (- 1.30 kg) and NDF (- 0.63 kg) were lower  
461 ( $P < 0.02$ ) in pregnant than in non-pregnant heifers at 267 DOP. In all collection  
462 periods, DM passage rate was greater ( $P < 0.09$ ) in pregnant than in non-pregnant  
463 heifers, and the difference was more evident at late gestation. Nitrogen balance and  
464 ruminal fermentation parameters were similar between PS. Pregnant heifers were  
465 more efficient ( $P \leq 0.09$ ) over time to synthesize microbial protein. Late-gestating beef  
466 heifers have faster digesta passage rate and are less efficient at extracting energy from  
467 feed compared to non-pregnant animals, changing the predicted feed total digestible  
468 nutrients.

469

470 **Keywords:** dry matter intake, gestation, intestinal digestibility, ruminal capacity,  
471 ruminal digestibility, ruminal retention time

472

## 473 1. Introduction

474 Properly estimating feed intake is essential to accurately meet the requirements  
475 for the maintenance and production of beef cattle (Allen, 1996). However, traditional  
476 models of feed intake regulation in ruminants normally do not include physical and  
477 physiological factors that affect feed intake of pregnant cows (NASEM, 2016).  
478 Particularities, such as the influence of fetal growth in reducing ruminal capacity, are  
479 difficult to model and are the main causes of the variations in voluntary intake observed  
480 at this physiological status (PS) of cattle (Ingvarsen, 1994). It is known that beef cows  
481 have a dramatic increase in energy and protein demand during late gestation (Gionbelli



482 et al., 2016), however their ruminal capacity is limited (Stanley et al., 1993). Although  
483 ruminal volume is reduced close to 40% at the end of gestation (Gionbelli, 2013), the  
484 reduction in feed intake is not proportional (Ingvarlsen et al., 1992), suggesting the  
485 development of compensation mechanisms to increase feed intake. These  
486 compensatory mechanisms may be related to the adjustment of feed digestibility and  
487 passage rate.

488 Digestibility and passage rate are inversely proportional; in other words,  
489 decreases in apparent total-tract digestibility of diet components may be explained in  
490 part by the increase in passage rate (Colucci et al., 1982; Ribeiro et al., 2015). The  
491 movement of digesta through the gastrointestinal tract of pregnant beef cows is  
492 increased in late gestation (Vanzant et al., 1991; Hanks et al., 1993; Stanley et al.,  
493 1993; Linden et al., 2014). Conversely, studies evaluating the effects of pregnancy on  
494 digestibility in cattle are not consistent, noticeably in zebu cattle. The *in vitro* dry matter  
495 (DM) digestibility in pregnant heifers was lower compared with non-pregnant heifers  
496 (Scheaffer et al., 2001). Vanzant et al. (1991) and Hanks et al. (1993) found no  
497 difference in organic matter (OM) and DM digestibility, respectively, in pregnant and  
498 non-pregnant animals. On the other hand, DM digestibility was greater for pregnant  
499 animals than for non-pregnant animals and decreased more overtime for non-pregnant  
500 animals than for pregnant animals (Linden et al., 2014). However, these results  
501 represent the total-tract apparent digestibility, which is the sum of ruminal and post-  
502 ruminal digestibility. Therefore, to our understanding, this is the first study evaluating,  
503 in pregnant and rumen cannulated zebu beef heifers, the changes in partial digestions  
504 (ruminal and intestinal) as a function of pregnancy time. Once digestion is altered,  
505 changes in ruminal fermentation parameters can be verified (Allen, 1997).

506 We hypothesized that as a way to compensate for the reduction in ruminal  
507 volume caused by the compression generated due to fetal growth, beef heifers  
508 increase the passage rate and reduce the ruminal digestibility of diet components  
509 without altering intestinal digestibility. The aim was to quantify the effects of  
510 physiological status (pregnant and non-pregnant) and pregnancy time on maternal  
511 performance, feed intake, digestibility and digestion kinetics in zebu beef heifers.

512

## 513 **2. Material and Methods**

514 The experiment was carried out in the Department of Animal Sciences Beef Unit  
515 of the Federal University of Lavras (UFLA) in Lavras, MG, Brazil. All procedures  
516 involving animal care and management were approved by the UFLA Ethics Committee  
517 on Animal Use (protocol number: 048/16).

518

### 519 *2.1. Animals, housing, and feeding*

520 Twelve rumen cannulated zebu beef heifers (body weight, BW =  $417 \pm 95.6$  kg)  
521 were used. Heifers were artificially inseminated following an ovarian synchronization  
522 protocol. Pregnancy was detected sixty days later via transrectal ultrasonography.  
523 Seven heifers were grouped as pregnant, and five non-pregnant heifers were used as  
524 controls. The control group was used to compare with pregnant heifers at different  
525 stages of gestation since the nutritional composition of the diet ingredients and climate  
526 conditions could vary throughout time. The heifers were group-housed in pastures with  
527 water and mineral supplement available up to 85 days of pregnancy (DOP). Then,  
528 heifers were allocated to individual pens (80 m<sup>2</sup> with 16 m<sup>2</sup> of covered area) until  
529 calving. A period of ten days was allowed for housing and diet adaptation, during which  
530 the DM offered was gradually increased until voluntary intake was reached.

531 All heifers received the same diet throughout the experiment. The experimental  
532 diet was based on medium-quality corn silage and concentrate supplement containing  
533 ground corn, soybean meal, urea plus ammonium sulfate, and mineral mixture (Table  
534 1). Experimental diets were formulated according to the Nutrient Requirements of Zebu  
535 and Crossbred Cattle - BR-CORTE 3.0 (Valadares Filho et al., 2016), to allow *ad*  
536 *libitum* intake without large accumulation of body reserves and an adequate  
537 maintenance of gestation. The use of 95.95% medium-quality corn silage sought to  
538 approach the nutritional quality of the diet to that of to grazing conditions plus a protein-  
539 based supplement. Heifers were fed twice a day at 0800 h and 1400 h. Daily orts were  
540 removed and weighed before the morning feeding. Free access to good-quality water  
541 was available.

542

## 543 2.2. *Collection periods and sampling*

544 Samples were obtained at six collection periods throughout gestation (107, 170,  
545 208, 240, 267, and 286 DOP). Collection periods 107, 208, and 267 DOP were  
546 performed for 11 straight days, while periods 170, 240, and 286 DOP for five  
547 consecutive days. The feed intake control started at 95 DOP until parturition. Corn  
548 silage samples were collected daily, and concentrate-ingredient samples were  
549 obtained immediately before mixing. Orts per animal were sampled daily during each  
550 collection period. Heifer BW was measured at the beginning and the end of each  
551 collection period, before the morning feeding. Body condition score (BCS) was  
552 assessed on day five of each collection period and was obtained by the average score  
553 of three trained observers using a 9-point scale (1 = emaciated; 9 = obese).

554 Feces spot collection was performed on days one to five of each collection  
555 period to determine the coefficient of the apparent total-tract digestibility of dietary

556 components. Fecal samples were collected at 10-h intervals within a day and at 16-h  
557 intervals between days (day one, 0600 and 1600 h; day two, 0800 and 1800 h; day  
558 three, 1000 and 2000 h; day four, 1200 and 2200 h; day five, 1400 h), totaling nine  
559 samples per collection period. Fecal samples were immediately frozen at  $-20^{\circ}\text{C}$  until  
560 further analyses. On the same days of fecal sampling, spot urine samples were  
561 collected to assess microbial crude protein (MCP) synthesis and the excretion of  
562 urinary nitrogenous compounds. Five-spot urine samples were obtained by stimulating  
563 the area below the vulva (day one, 0600 h; day two, 1800 h; day three, 1000 h; day  
564 four, 2200 h; day five, 1400 h). Urine samples were filtered, and a 12 mL aliquot was  
565 immediately acidified by diluting one volume of urine with four volumes of sulfuric acid  
566 ( $\text{H}_2\text{SO}_4$ ) at 0.036 mol/L to avoid N loss. Then, samples were frozen at  $-20^{\circ}\text{C}$  for further  
567 analysis.

568         During the 107, 208, and 267 DOP collection periods, samplings of omasal and  
569 ruminal digesta were performed. Omasal digesta sampling was performed according  
570 to Huhtanen et al. (1997), with modifications described by Leão (2002). Omasal  
571 digesta was collected twice a day, at 12-h intervals within a day, and at 16-h intervals  
572 between days to avoid possible variation in digesta flux related to collection time.  
573 Samples are collected at 0600 and 1800 h on day six, at 1000 and 2200 h on day  
574 seven, and 1400 and 0200 h of the next day on day eight, totaling six samples per  
575 collection period. A sample of approximately 600 mL of digesta per animal was  
576 collected and frozen at  $-20^{\circ}\text{C}$  for further analysis. For determination of ruminal DM  
577 outflow, two indicators were utilized: Co-EDTA as the fluid phase and small particles  
578 indicator (Udén et al., 1980) and indigestible neutral detergent fiber (iNDF) as the solid-  
579 phase indicator. The Co-EDTA was wrapped in 1.5 g paper cartridges, and a total of  
580 six g was provided daily, administered four times in 6-h intervals (0600, 1200, 1800,

581 and 0000 h), directly through the ruminal cannula. The administration of Co-EDTA  
582 started three days before the first omasal digesta collection until the last collection.

583 To determine the ruminal pools, rates of intake (ki), passage (kp), and digestion  
584 (kd) of diet components, on day nine, complete evacuation of the rumen was performed  
585 approximately four hours after feeding, according to procedures described by Allen  
586 and Linton (Allen and Linton, 2007). After evacuation, the total digesta was weighed  
587 and then filtered through four layers of cheesecloth to separate the solid and fluid  
588 phases, which were weighed and sampled for further analysis. Then, the digesta was  
589 put back into the respective rumen. On day 11, ruminal evacuation was also performed  
590 one hour before feeding, theoretically the time the rumen is at its least volume.

591 To evaluate the pH, volatile fatty acids (VFA), and concentration of ruminal  
592 ammoniacal N (N-NH<sub>3</sub>), ruminal fluid was sampled on collection periods 107, 208, and  
593 267 DOP. Samples were collected manually from the ventral sections of the rumen  
594 before (time 0) and four hours after (time 4) morning feeding. Ruminal fluid samples  
595 were filtered through a triple layer of gauze, and 50 mL of the ruminal liquid were used  
596 immediately to determine pH, in a pH-meter (model HI 2221, Hanna Instruments,  
597 Woonsocket, RI, USA). At the same time, 20 mL aliquots of filtered ruminal fluid were  
598 shocked frozen in liquid N to inhibit microbial growth. Then, samples were frozen at -  
599 20 °C for VFA analysis. Additionally, 1 mL of H<sub>2</sub>SO<sub>4</sub> diluted in distilled water (1:1) was  
600 added to 50 mL aliquots of filtered ruminal fluid and frozen at -20 °C for N-NH<sub>3</sub> analysis.

601

### 602 2.3. *Laboratory procedures, analyses, and calculations*

603 Heifer BW was determined by the average between the BWs at the beginning  
604 and end of each collection period. Pregnancy was considered as an extra component  
605 of the heifer (referred mathematically as pregnancy component, PREG). The PREG

606 allows the estimating portion of the BW of a pregnant heifer that is a function of  
607 pregnancy. The PREG includes all tissues that increase due to the pregnancy and is  
608 equal to the gravid uterus plus udder accretion during pregnancy. The PREG was  
609 estimated using DOP, heifer BW and BCS, and calf BW at calving (Gionbelli et al.,  
610 2015). The BW of a pregnant heifer minus PREG or the BW of a non-pregnant heifer  
611 was called BWnp. To reduce the fill effect and improve the accuracy of measurements,  
612 BW was used to estimate the shrunk body weight (SBW), as follows:  $SBWnp = 0.8084$   
613  $\times BWnp^{1.0303}$  (Gionbelli et al., 2015). The SBW of a pregnant heifer minus PREG or the  
614 SBW of a non-pregnant heifer was called SBWnp.

615 At the end of each collection period, omasal digesta were thawed at room  
616 temperature, and an animal composite sample was obtained. Two-thirds of composite  
617 samples were filtered through a 100- $\mu$ m nylon mesh filter with a 44% open area (Sefar  
618 Nitex 100/44; Sefar, Thal, Switzerland), resulting in two phases. The portion retained  
619 in the mesh was designated as particle phase, and the filtered portion was named fluid  
620 + small particle phase. Thawed samples of corn silage, Orts, feces, ruminal and omasal  
621 digesta were pre-dried in a forced-air oven at 65°C for 72 h. Then, samples were  
622 ground in a knife-mill with a 2-mm sieve for iNDF, and with a 1-mm sieve for DM, OM,  
623 crude protein (CP), ash- and protein-free neutral detergent fiber (apNDF), and ether  
624 extract (EE).

625 Corn silage samples were composed weekly or by collection period.  
626 Furthermore, composite samples of Orts, feces, ruminal digesta for each heifer, and  
627 collection period were obtained. A composite sample of ruminal digesta was obtained  
628 from the dry samples of the solid and fluid phases of the two ruminal evacuations (one  
629 hour before and four hours after feeding), based on the dry weight of each sample.

630 Ground samples were analyzed for DM (method INCT-CA G-003/1), OM  
631 determined by ash (method INCT-CA M-001/1), and CP obtained by total N, using the  
632 micro-Kjeldahl technique (method INCT-CA N-001/1). Additionally, the EE was  
633 determined after petroleum ether extraction (method INCT-CA G-005/1) and iNDF  
634 (INCT-CA F-009/1), according to Detmann et al. (2012). Ash- and protein-free NDF  
635 were analyzed using filtration in porous crucibles with heat-stable  $\alpha$ -amylase and  
636 sodium sulfite (Van Soest et al., 1991). Non-fiber carbohydrates (NFC) were calculated  
637 according to Detmann and Valadares Filho (Detmann and Valadares Filho, 2010),  
638 where  $\text{NFC (\% of DM)} = 100 - [\text{CP} - (\text{CP derived from urea} + \text{urea}) + \text{NDF} + \text{EE} +$   
639  $\text{ash}]$ . Cobalt content in omasal digesta was analyzed in a mineral solution prepared  
640 according to method INCT-CA M-004/1 (Detmann et al., 2012) using an atomic  
641 absorption spectrophotometer (model AA-7000; Shimadzu Corp., Kyoto, Japan).

642 The apparent total-tract digestibilities of DM, OM, CP, apFDN, and NFC were  
643 determined by the difference between intake and the fecal content divided by the  
644 intake. The content of total digestible nutrients (TDN) was obtained according to  
645 recommendations from the NRC (2001).

646 Ruminal DM and apNDF outflows were estimated by the double-indicator (Co-  
647 EDTA and iNDF) system (France and Siddons, 1986). For the double-indicator system  
648 calculation, concentrations of indicators in different digesta phases were used to  
649 calculate the reconstitution factor. The reconstitution factor indicates the units of the  
650 digesta phase that must be removed (when negative) or added (when positive) to non-  
651 representative digesta to reconstitute the true digesta (France and Siddons, 1986).

652 The composite samples of ruminal digesta were used to calculate the ruminal  
653 pool of DM and apNDF. Rates of intake, passage, and digestion were estimated using  
654 the pool-and-flux method (Robinson et al., 1987). The  $k_i$  was calculated by dividing the

655 diet component intake by its respective ruminal pool size. The  $k_p$  was obtained by  
656 dividing the ruminal outflow of the diet component by its respective ruminal pool size.  
657 The  $k_d$  was calculated as  $k_i$  minus  $k_p$ .

658 Urine samples were thawed, and a composite for each heifer and collection  
659 period was performed. Urine total N was determined using the micro-Kjeldahl  
660 technique [method INCT-CA N-001/1; (Detmann et al., 2012)]. Urine creatinine  
661 concentration was analyzed using a commercial kit (Creatinine K016, Bioclin, Belo  
662 Horizonte, Brazil). Allantoin concentration was quantified according to the colorimetric  
663 method (Chen and Gomes, 1992). Uric acid was estimated using allantoin  
664 concentration (Santos et al., 2016), as follows: uric acid (mmol/d) =  $0.1104 \times$  allantoin  
665 (mmol/d). Microbial CP synthesis was estimated by using the technique of the purine  
666 derivatives in urine (Chen and Gomes, 1992). Urine volume was estimated using  
667 creatinine concentration as a marker and assuming a daily creatinine excretion (mg/d)  
668 of  $37.88 \times SBW^{0.9316}$ , where SBW is the shrunk body weight in kg (Santos et al., 2016).  
669 Excretion of purine derivatives in urine was calculated by the sum of the allantoin and  
670 uric acid excretions, which were obtained by the product between their concentrations  
671 in urine by the daily urinary volume. Absorbed purines were calculated from the  
672 excretion of purine derivatives (Prates et al., 2012), as follows: absorbed purines  
673 (mmol/d) = excretion of purine derivatives (mmol/d) –  $(0.389 \times BW^{0.75})/0.99$ ; where 0.99  
674 = recovered absorbed purines. The  $0.389 \times BW^{0.75}$  value = endogenous excretion of  
675 purine derivatives (mmol/d). Ruminal MCP synthesis was calculated as a function of the  
676 absorbed purines (Prates et al., 2012), as follows: MCP =  $70 \times$  absorbed purines  
677 (mmol/d)/ $(0.93 \times 0.11 \times 1000)$ ; where 70 = purine N content (mg N/mol), 0.93 = purine  
678 digestibility and 0.11 = relation of purine N:total N of microorganisms. The efficiency of  
679 MCP synthesis was calculated dividing the amount of MCP by intake of CP, digestible



680 OM, and TDN. The total-tract N balance was calculated subtracting from the N intake  
681 the amount excreted via urine and feces. Ruminal N balance was considered the  
682 difference between N intake and ruminal outflow of N.

683 Ruminal fluids for VFA analyses were thawed, filtered through sterile syringe  
684 filter with 0.45  $\mu\text{m}$  pore sizes, centrifuged at 1,000  $\times$  g for 30 min at 4°C, and acidified  
685 with 0.2 mL of  $\text{H}_2\text{SO}_4$  at 0.1 mol/L. Volatile fatty acid concentrations were determined  
686 using a gas chromatograph (Varian Model CP3800; Varian, Inc, Walnut Creek, CA)  
687 equipped with a Nukol capillary GC column (length: 15 m; inner diameter: 0,53 mm;  
688 stationary phase film thickness: 0,50  $\mu\text{m}$ ) bonded phase - acid modified polyethylene  
689 glycol [Supelco, Bellefonte, PA]), where  $\text{N}_2$  was used as a carrier gas at a flow rate of  
690 2.5 mL/min. The following temperature programming was used in the oven: 110°C for  
691 1 min followed by a ramp of 6°C/min to 160°C and ramp of 30°C/min up to 195°C a  
692 with a final plateau of 5 min. The injection port and detector port temperatures were  
693 220 and 250°C, respectively. Ruminal  $\text{N-NH}_3$  concentrations were determined  
694 according to method INCT-CA N-006/1 described by Detmann et al., 2012.

695

#### 696 2.4. *Statistical analyses*

697 Data were analyzed through the mixed model methodology (procedure MIXED  
698 of SAS 9.2, SAS Inst. Inc., Cary, NC), considering the PS effect (pregnant or non-  
699 pregnant) and the DOP as fixed effects and animal as the random effect. When  
700 appropriate, initial BW was included as a covariate in the model. Once repeated  
701 measurements were taken from the same animal (for DOP), the subject animal nested  
702 within treatment was included in the repeated measurement statement. For every  
703 DOP, the PS effect on the measured variable was estimated using the “estimate  
704 statement” of SAS. The  $P$ -value  $< 0.10$  was adopted as a critical level of probability for

705 the occurrence of Type I error. Tendency was determined as  $0.15 > P \geq 0.10$ . Except  
706 for body weight variables, the isolated effect of DOP is not discussed, as it is related  
707 to environmental and dietary and environmental variations over time and not exactly  
708 to the study factor.

709

### 710 **3. Results**

711

#### 712 **3.1. Body weight**

713 Mean pregnancy length and calf BW at birth were  $292 \pm 4$  days and  $31.3 \pm 6.7$   
714 kg, respectively. Initial BW and BCS did not differ ( $P \geq 0.80$ ) between pregnant and  
715 non-pregnant heifers (Table 2). Moreover, BW, SBW, and BCS also did not vary ( $P \geq$   
716  $0.29$ ) between PS. However, BW and SBW increased ( $P < 0.01$ ) over days in both  
717 groups. On the other hand, BWnp and SBWnp decreased ( $P = 0.01$ ) in pregnant  
718 compared with non-pregnant heifers over days (Fig. 1A and 1B). In pregnant heifers  
719 BWnp and SBWnp were lower ( $P < 0.01$ ) than in non-pregnant heifers at late gestation  
720 (267 and 286 DOP). Whereas, at 107, 170, 208, and 240 DOP, BWnp and SBWnp  
721 were similar ( $P = 0.27$ ) between pregnant and non-pregnant heifers. The estimated  
722 PREG increased over DOP (Fig. 1C).

723

#### 724 **3.2. Feed intake**

725 There was an interaction ( $P \leq 0.08$ ) between the effects of DOP and PS on DM,  
726 OM, CP, apNDF, iNDF, and TDN intake (kg/d) such that these parameters decreased  
727 in pregnant heifers over gestation time, but not in non-pregnant heifers. (Table 3). At  
728 107 DOP, TDN intake was greater ( $P = 0.05$ ), while at 286 DOP, TDN intake tended  
729 to be lower ( $P = 0.10$ ) in pregnant compared to non-pregnant heifers. Furthermore, at

730 286 DOP, OM and apNDF intake were lower ( $P = 0.09$ ), and DM, CP, and iNDF intake  
731 tended to be lower ( $P = 0.11$ ) in pregnant compared to non-pregnant heifers. Likewise,  
732 when expressed as grams per kilogram of BW, DM (Fig. 2), apNDF, iNDF, and TDN  
733 (Fig. 3) intakes also were affected by an interaction effect ( $P \leq 0.05$ ). At 107 DOP, TDN  
734 intake was greater ( $P = 0.04$ ) and DM, apNDF, and iNDF intake tended to be greater  
735 ( $P \leq 0.13$ ) in pregnant compared to non-pregnant heifers, whereas, at 286 DOP, these  
736 same intakes were lower ( $P \leq 0.09$ ) in pregnant heifers.

737

### 738 3.3. *Apparent total-tract, ruminal, and intestinal digestibility*

739 The apparent total-tract digestibility (g/kg of DM) of DM and apNDF were lower  
740 ( $P \leq 0.09$ ) and OM digestibility tended to be lower ( $P = 0.11$ ) in pregnant compared to  
741 non-pregnant heifers (Table 4). There was an interaction effect ( $P = 0.04$ ) between PS  
742 and DOP on CP apparent total-tract digestibility. Pregnant heifers had similar ( $P \geq 0.22$ )  
743 CP apparent total-tract digestibility until 240 DOP, but greater ( $P < 0.01$ ) digestibility at  
744 267 and 286 DOP compared with non-pregnant heifers (Fig. 4). In contrast, the ruminal  
745 digestibility (g/kg of DM) of OM and apNDF was lower ( $P \leq 0.09$ ) in pregnant compared  
746 to non-pregnant heifers, but the PS did not affect ( $P \geq 0.22$ ) the DM, CP or NFC ruminal  
747 digestibility. Regarding intestinal digestibility (g/kg of the amount reaching the  
748 omasum), no difference was observed in the diet components depending on PS.

749

### 750 3.4. *Ruminal pool and passage rate*

751 There was an interaction between PS and DOP ( $P = 0.01$ ) on the ruminal WM  
752 pool (kg; Table 5). In pregnant heifers the ruminal WM pool at 107 and 208 DOP were  
753 similar ( $P \geq 0.30$ ), but was lower ( $P < 0.01$ ) at 267 DOP than in non-pregnant heifers  
754 (Fig. 5). Similarly, there was a PS  $\times$  DOP interaction effect ( $P \leq 0.02$ ) on the ruminal

755 pool of DM and apNDF. In pregnant heifers the ruminal pool of DM (- 1.30 kg) and  
756 apNDF (- 0.63 kg) were lower ( $P < 0.01$ ) than in non-pregnant heifers at 267 DOP.  
757 Conversely, the ruminal outflow of DM and apNDF were greater ( $P \leq 0.04$ ) at 107 and  
758 208 DOP (average of 0.74 and 0.40 kg, respectively) in pregnant heifers. Additionally,  
759 an interaction effect ( $P \leq 0.09$ ) between PS and DOP was found on DM kp and ki and  
760 apNDF ki and kd (Table 5). Pregnant heifers had greater DM kp (Fig. 6) and ki in all  
761 collection periods; however, the difference was numerically greater at 267 DOP.  
762 Concerning apNDF ki and kd, at 267 DOP, pregnant compared to non-pregnant heifers  
763 increased by 0.85 %/h and 0.30 %/h, respectively. Also, pregnant heifers had greater  
764 ( $P \leq 0.05$ ) DM kd and kp apNDP, and lower ( $P < 0.01$ ) retention time for DM and apNDF  
765 than non-pregnant heifers.

766

### 767 3.5. *Nitrogen balance and efficiency of microbial protein synthesis*

768 No difference was found between PS on N balance, with the exception of N  
769 ruminal outflow (Table 6). An interaction between the effect of PS and DOP was  
770 detected ( $P = 0.03$ ) on N ruminal outflow (g/day), but this effect was likely a  
771 consequence of variation over time than PS. There was an interaction effect ( $P \leq 0.09$ )  
772 between PS and DOP on MCP expressed per unit of CP, digestible OM (DOM), and  
773 TDN intakes (Fig. 7). Pregnant heifers were more efficient over pregnancy time,  
774 achieving greater ( $P \leq 0.06$ ) difference at 286 DOP.

775

### 776 3.6. *Ruminal fermentation parameters*

777 Before morning feeding, there was no difference between PS in ruminal  
778 fermentation parameters (Table 7). On the other hand, four hours after the morning  
779 feeding, total volatile fatty acids concentration (mmol/L) tended to be greater ( $P = 0.12$ )

780 in pregnant heifers. Additionally, there was a tendency ( $P = 0.14$ ) for an interaction  
781 effect between PS and DOP on the propionic acid concentration (mmol/L) at four hours  
782 after the morning feeding. At late gestation (267 DOP), the concentration of propionic  
783 acid in the ruminal fluid were 15.1 mmol/L and 19.1 mmol/L in pregnant and non-  
784 pregnant heifers, respectively.

785

## 786 **4. Discussion**

### 787 *4.1. Body weight*

788 During pregnancy, BW increase was probably due to the deposition of body  
789 tissue reserves or due to the growth of pregnancy-related components, such as the  
790 gravid uterus and mammary gland (Gionbelli et al., 2015). Rendering the use of BW to  
791 assess beef cow pregnancy performance unsuitable. In this trial, when estimating  
792 pregnancy components, a loss of heifer maternal body reserve was detected at late  
793 gestation. This observation indicates the mobilization of maternal stores to support the  
794 growing fetus and developing mammary gland requirements (Scheaffer et al., 2001;  
795 Linden et al., 2014). Insufficient DM intake and low-quality diet may have led to such  
796 mobilization.

797 Another pattern, commonly observed during pregnancy, is the transition from  
798 an anabolic to a catabolic state as pregnancy progresses (Robinson, 1986). Results  
799 indicate that this transition in maternal metabolism occurs after 240 days of gestation,  
800 corroborating results of previous studies (Scheaffer et al., 2001; Meyer et al., 2010).

801

### 802 *4.2. Feed intake*

803 The current study showed that pregnant heifers had a greater TDN intake at the  
804 end of the second third of gestation (170 DOP), suggesting animal adaptation to body

805 reserve accumulation to sustain late pregnancy high metabolic demand. Similarly,  
806 voluntary DM intake in pregnant dairy heifers reached a peak between 15 and 17  
807 weeks before calving (Ingvarlsen and Andersen, 2000), i.e., at the end of the second  
808 third of gestation. Indeed, in this trial, the accumulation of maternal tissues occurred at  
809 the same stage of pregnancy. Additionally, the greater intake at this period may be the  
810 result of a faster DM kp observed in pregnant heifers.

811 On the other hand, DM and TDN intakes decreased throughout pregnancy,  
812 coincidentally with a lower ruminal pool of wet matter (WM), DM, and apNDF, which are  
813 indicative of low ruminal capacity. These results agree with studies that showed a  
814 reduction in feed intake and ruminal fill at late pregnancy (Hanks et al., 1993; Stanley  
815 et al., 1993; Scheaffer et al., 2001). These observations provide sufficient evidence to  
816 support a physical effect of pregnancy in reducing ruminant DM intake at late gestation.  
817 However, it is unlikely that the decrease in ruminal volume is the only cause of the late  
818 gestation intake decline. It is important to note that the effects of physical compression  
819 coincide with the changes in endocrine factors and body reserves, mediated in  
820 response to the advancement of gestation and preparation for future lactation (Forbes,  
821 2013).

822 Nevertheless, the reduction observed in intake was not proportional to the  
823 reduction in the ruminal pool of DM at late gestation. While the ruminal pool was  
824 reduced to around 23%, the intake reduction was only 11%. Consequently, this  
825 disproportionality caused an increase in DM ki for pregnant heifers in the present study,  
826 considering that DM ki is a ratio between DM intake and ruminal pool (Robinson et al.,  
827 1987).

828

829 4.3. *Passage rate*

830           The movement of digesta through the rumen of pregnant beef heifers was  
831 increased probably to compensate for the loss of ruminal capacity caused by fetal  
832 growth in late gestation. Passage rates are affected by a wide variety of factors exerting  
833 different effects, including animal and feed factors (Moyo and Nsahlai, 2018). Okine  
834 and Mathison (Okine and Mathison, 1991) indicated that ruminal kp increased  
835 concomitantly with intake. But, during late gestation, feed intake seems to be not the  
836 causative factor for changes in kp. Pregnant cows and heifers fed for *ad libitum* intake  
837 had faster digesta kp than their non-pregnant counterparts (Linden et al., 2014).  
838 Likewise, ruminal indigestible ADF kp increased from 61 days to six days prepartum in  
839 mature beef cows fed *ad libitum* (Stanley et al., 1993).

840           On the other hand, Hanks et al. (Hanks et al., 1993) found that limit-fed pregnant  
841 cows had increased particulate kp and decreased ruminal and total-tract mean  
842 retention time when compared to non-pregnant cows. Thus, an increase in kp occurs  
843 even when intake does not change. It is concluded that the impact of pregnancy on the  
844 digesta kp occurs through factors other than feed intake. Nevertheless, the reasons for  
845 the increase in the kp are still unclear. Neural or hormonal factors might contribute to  
846 an increase in kp (Stanley et al., 1993).

847           Changes in digesta passage from the ruminoreticulum are associated with the  
848 strength (Al-Shboul et al., 2019) and duration (Okine and Mathison, 1991) of reticular  
849 contraction. In this context, the frequency of primary ruminoreticular movements  
850 increased as the pregnancy progressed in pregnant sheep (Stafford, 1991); however,  
851 no studies were found evaluating ruminoreticular motility during pregnancy of cattle. In  
852 general, the ruminoreticular motility is controlled by smooth muscle contractility, which  
853 in turn is regulated by extrinsic (sympathetic and parasympathetic neurons) and  
854 intrinsic (sensory and motor neurons) factors of the enteric system and specific

855 hormones (Costa et al., 2000). Several hormones are linked to this motility function,  
856 such as secretin, peptide YY, neurotensin, gastrin, gastrin-releasing peptide,  
857 cholecystokinin, somatostatin, ghrelin, and motilin (Kitazawa and Kaiya, 2019).  
858 Recently, it was verified that total ghrelin concentration increases during the last  
859 months of pregnancy in adult cows but not in heifers (Chouzouris et al., 2018).  
860 According to Forbes (1986), circulating estradiol concentrations may increase kp.  
861 However, recent studies, in humans, have shown that estradiol is responsible for  
862 relaxing the smooth muscles of the stomach and other organs (Al-Shboul et al., 2018;  
863 Al-Shboul et al., 2019). Thus, although estradiol concentrations are higher in pregnant  
864 cows (Hanks et al., 1993), they are unlikely to be the cause of the increase in kp.

865 The motility of ruminant digestive tracts can also be controlled by tension  
866 receptors (located in the muscular wall) and epithelial receptors in the reticulorumen  
867 (Forbes and Barrio, 1992). Therefore, the lower ruminal volume of pregnant heifers  
868 can increase the mechanical stimulation caused by digesta on these receptors and  
869 can also increase ruminoreticular motility and, consequently, digesta passage rate.

870

#### 871 4.4. *Digestibility*

872 The ruminal digestibility of apNDF and OM was impaired in pregnant heifers  
873 corroborating our hypothesis. To our knowledge, this is the first study that reports  
874 changes in the ruminal digestibility of beef cattle during pregnancy. The faster ruminal  
875 kp leads to lower digestibility due to the shorter time feed is exposed to digestive  
876 processes (Ribeiro et al., 2015). Indeed, our results showed that pregnant heifers had  
877 a faster passage of ruminal DM and apNDF and shorter retention time of these diet  
878 components in the rumen, mainly at late gestation. As a consequence of lower ruminal  
879 digestibility, the apparent total-tract digestibility of apNDF and DM was lower, and OM



880 tended to be lower in pregnant heifers since these components had no alteration in  
881 intestinal digestibility.

882 In contrast, it was observed that the CP apparent total-tract digestibility  
883 increased in pregnant heifers with the course of gestation. At the same time, pregnant  
884 heifers had greater efficiency of MCP synthesis. Therefore, assuming that MCP has  
885 high intestinal digestibility, around 80% (Mariz et al., 2018), the greater CP digestibility  
886 can be determined by the better quality of CP reaching the intestines.

887

#### 888 4.5. *Nitrogen balance*

889 In grazing bovines, the excretion of N in urine is linearly related to N intake  
890 (Hoekstra et al., 2007). Despite the lower values of N intake in pregnant heifers at late  
891 gestation, the N excretion did not change. Nitrogen balance is correlated to energy  
892 balance in pregnant ruminants (Bauman and Currie, 1980). The conceptus has a high  
893 demand for glucose and amino acids at the end of pregnancy (Bell et al., 2005). Thus,  
894 although the lower N intake at late gestation, the catabolism of amino acids to support  
895 the demands of fetal energy possibly increased the excretion of nitrogen compounds  
896 (Rotta et al., 2015).

897

#### 898 4.6. *Efficiency of microbial crude protein synthesis*

899 The efficiency of MCP synthesis is strongly correlated to DM intake (Broderick  
900 et al., 2010). Furthermore, MCP synthesis is increased by a longer digesta retention  
901 time in the rumen, or a reduced digesta kp (Huhtanen et al., 2016). For these reasons,  
902 it was expected that during late gestation, pregnant heifers would have a low efficiency  
903 in synthesizing MCP. This hypothesis, however, was not confirmed. Actually, the  
904 efficiency of MCP synthesis was greater in pregnant than in non-pregnant heifers

905 because even though the intake of pregnant heifers was lower, MCP synthesis was  
906 similar between PS. The MCP synthesis was estimated from the purine derivatives  
907 content in the urine (Chen and Gomes, 1992). In turn, urinary volume was estimated  
908 by the urinary creatinine concentration. Creatinine is formed from muscle metabolism  
909 and is excreted at a constant rate relative to muscle mass and, consequently, body  
910 weight (Costa e Silva et al., 2012). There is evidence that creatinine excretion changes  
911 depending on pregnancy status or pregnancy time (Hare et al., 2019; Whittet et al.,  
912 2019). However, more studies need to be carried out to corroborate such a hypothesis,  
913 especially in zebu cows raised in tropical conditions. Pregnant heifers underwent  
914 intense catabolism of skeletal muscle tissue at late pregnancy due to low protein intake  
915 and high requirements of gestation. Thus, this estimated microbial yield may not be  
916 representative of the real MCP synthesis since total collection of urine was not  
917 performed.

918

#### 919 4.7. *Ruminal fermentation parameters*

920 Ruminal pH depends on saliva production, the balance of volatile fatty acids  
921 (VFA), the type and level of feed intake, and the exchange of bicarbonates and  
922 phosphates through the ruminal epithelium (Aschenbach et al., 2011). Whereas,  
923 rumen VFA concentration reflects the balance between production and clearance by  
924 passage with the fluid phase into the omasum or by absorption through the ruminal  
925 wall (Lopez et al., 2003). Although pregnant heifers have lower intake and digestibility  
926 and a higher DM passage rate, ruminal pH did not change and the VFA concentration  
927 in the rumen had only a neglectable change. Additionally, N-NH<sub>3</sub> is a very potent buffer  
928 once NH<sub>3</sub> can immediately bind H<sup>+</sup> to form NH<sub>4</sub><sup>+</sup> in the ruminal content. Ruminal N-  
929 NH<sub>3</sub> removal occurs through the use of ruminal bacteria, efflux to the omasum, or

930 absorption across the ruminal wall (Aschenbach et al., 2011). Similarly, to pH and VFA,  
931 N-NH<sub>3</sub> concentration was not different between PS.

932

## 933 **5. Conclusions**

934 As pregnancy progresses, ruminal capacity is reduced leading to lower feed  
935 intake and an increase of the digesta passage rate, impairing the ruminal digestibility  
936 of dietary dry matter and fiber. Thus, late-gestating beef heifers are less efficient at  
937 extracting energy from feed compared to non-pregnant animals, changing the feed  
938 predicted total digestible nutrients.

939

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949

## 950 **References**

951 Al-Shboul, O.A., Al-Rshoud, H.J., Al-Dwairi, A.N., Alqudah, M.A., Alfaqih, M.A.,  
952 Mustafa, A.G., Jaafar, M., 2019. Changes in Gastric Smooth Muscle Cell  
953 Contraction during Pregnancy: Effect of Estrogen. J. Pregnancy 2019, 4302309.

- 954 Al-Shboul, O.A., Nazzal, M.S., Mustafa, A.G., Al-Dwairi, A.N., Alqudah, M.A., Abu  
955 Omar, A., Alfaqih, M.A., Alsalem, M.I., 2018. Estrogen relaxes gastric muscle  
956 cells via a nitric oxide- and cyclic guanosine monophosphate-dependent  
957 mechanism: A sex-associated differential effect. *Exp. Ther. Med.* 16, 1685-  
958 1692.
- 959 Allen, M.S., Linton, J.V., 2007. In vivo methods to measure digestibility and digestion  
960 kinetics of feed fractions in the rumen. In: *Proc of 1st Simpósio Internacional*  
961 *Avanços em Técnicas de Pesquisa em Nutrição de Ruminantes*. Pirassununga,  
962 Brazil: Universidade de São Paulo, 72-89.
- 963 Allen, M.S., 1996. Physical constraints on voluntary intake of forages by ruminants. *J.*  
964 *Anim. Sci.* 74, 3063-3075.
- 965 Allen, M.S., 1997. Relationship between fermentation acid production in the rumen and  
966 the requirement for physically effective fiber. *J. Dairy Sci.* 80, 1447-1462.
- 967 Aschenbach, J.R., Penner, G.B., Stumpff, F., Gabel, G., 2011. Ruminant Nutrition  
968 Symposium: Role of fermentation acid absorption in the regulation of ruminal  
969 pH. *J. Anim. Sci.* 89, 1092-1107.
- 970 Bauman, D.E., Currie, W.B., 1980. Partitioning of nutrients during pregnancy and  
971 lactation: a review of mechanisms involving homeostasis and homeorhesis. *J.*  
972 *Dairy Sci.* 63, 1514-1529.
- 973 Bell, A., Greenwood, P., Ehrhardt, R., 2005. Regulation of metabolism and growth  
974 during prenatal life. *Biology of growing animals*. Elsevier, pp. 3-34.
- 975 Broderick, G., Huhtanen, P., Ahvenjärvi, S., Reynal, S., Shingfield, K., 2010.  
976 Quantifying ruminal nitrogen metabolism using the omasal sampling technique  
977 in cattle - A meta-analysis. *J. Dairy Sci.* 93, 3216-3230.

- 978 Chen, X.B., Gomes, M., 1992. Estimation of microbial protein supply to sheep and  
979 cattle based on urinary excretion of purine derivatives: an overview of the  
980 technical details. Rowett Research Institute.
- 981 Chouzouris, T.M., Dovolou, E., Georgoulas, P., Rekkas, A., Dafopoulos, K.,  
982 Athanasiou, L., Fthenakis, G.C., Amiridis, G.S., 2018. Effects of pregnancy and  
983 short-lasting acute feed restriction on total ghrelin concentration and metabolic  
984 parameters in dairy cattle. *Theriogenology* 106, 141-148.
- 985 Colucci, P., Chase, L., Van Soest, P., 1982. Feed intake, apparent diet digestibility,  
986 and rate of particulate passage in dairy cattle. *J. Dairy Sci.* 65, 1445-1456.
- 987 Costa e Silva, L.F., Valadares Filho, S.d.C., Chizzotti, M.L., Rotta, P.P., Prados, L.F.,  
988 Valadares, R.F.D., Zanetti, D., Braga, J.M.d.S., 2012. Creatinine excretion and  
989 relationship with body weight of Nellore cattle. *Rev. Bras. Zootecn.* 41, 807-810.
- 990 Costa, M., Brookes, S.J., Hennig, G.W., 2000. Anatomy and physiology of the enteric  
991 nervous system. *Gut* 47, iv15-iv19.
- 992 Detmann, E., Souza, M., Valadares Filho, S.C., Queiroz, A., Berchielli, T., Saliba, E.O.,  
993 Cabral, L.S., Pina, D.S., Ladeira, M., Azevedo, J., 2012. Métodos para análise  
994 de alimentos. Visconde do Rio Branco: Suprema 214.
- 995 Detmann, E., Valadares Filho, S.C., 2010. On the estimation of non-fibrous  
996 carbohydrates in feeds and diets. *Arq. Bras. Med. Vet. Zoo.* 62, 980-984.
- 997 Forbes, J.M., 1986. The effects of sex hormones, pregnancy and lactation on  
998 digestion, metabolism and voluntary food intake. In: L. P. Milligan, W. L.  
999 Grovum, and A. Dobson (Ed.) *Control of Digestion and Metabolism in*  
1000 *Ruminants.* pp 420-435. Reston Publishing, Reston, VA.
- 1001 Forbes, J.M., Barrio, J., 1992. Abdominal chemo-and mechanosensitivity in ruminants  
1002 and its role in the control of food intake. *Exp. Physiol.* 77, 27-50.

- 1003 Forbes, J.M., 2013. The voluntary food intake of farm animals. Butterworth-  
1004 Heinemann.
- 1005 France, J., Siddons, R., 1986. Determination of digesta flow by continuous market  
1006 infusion. *J. Theor. Biol.* 121, 105-119.
- 1007 Gionbelli, M.P., 2013. Nutrient requirements and quantitative aspects of growth,  
1008 development and digestion of pregnant and non-pregnant Nellore cows  
1009 (Doctoral dissertation). Federal University of Viçosa, Viçosa, MG, Brazil..
- 1010 Gionbelli, M.P., Valadares Filho, S.C., Duarte, M.S. 2016. Nutritional requirements for  
1011 pregnant and non-pregnant beef cows, in: Valadares Filho, S.C. et al. (Eds.),  
1012 Nutrient Requirements of Zebu and Crossbred Cattle - BR-CORTE 3.0.  
1013 Suprema, Viçosa, Brazil, pp. 251-272.
- 1014 Gionbelli, M.P., Duarte, M.S., Valadares Filho, S.C., Detmann, E., Chizzotti, M.L.,  
1015 Rodrigues, F.C., Zanetti, D., Gionbelli, T.R., Machado, M.G., 2015. Achieving  
1016 body weight adjustments for feeding status and pregnant or non-pregnant  
1017 condition in beef cows. *PLoS One* 10, e0112111.
- 1018 Hanks, D.R., Judkins, M.B., McCracken, B.A., Holcombe, D.W., Krysl, L.J., Park, K.K.,  
1019 1993. Effects of pregnancy on digesta kinetics and ruminal fermentation in beef  
1020 cows. *J. Anim. Sci.* 71, 2809-2814.
- 1021 Hare, K.S., Wood, K.M., Acton, K., Fitzsimmons, C., Penner, G.B., 2019.  
1022 Oversupplying metabolizable protein in late gestation for beef cattle: effects on  
1023 prepartum BW, ruminal fermentation, nitrogen balance, and skeletal muscle  
1024 catabolism. *J. Anim. Sci.* 97, 407-423.
- 1025 Hoekstra, N., Schulte, R., Struik, P., Lantinga, E., 2007. Pathways to improving the N  
1026 efficiency of grazing bovines. *Eur. J. Agron.* 26, 363-374.

- 1027 Huhtanen, P., Brotz, P.G., Satter, L.D., 1997. Omasal sampling technique for  
1028 assessing fermentative digestion in the forestomach of dairy cows. *J. Anim. Sci.*  
1029 75, 1380-1392.
- 1030 Huhtanen, P., Ramin, M., Cabezas-Garcia, E.H., 2016. Effects of ruminal digesta  
1031 retention time on methane emissions: a modelling approach. *Anim. Prod. Sci.*  
1032 56.
- 1033 Ingvarsen, K.L., Andersen, J., 2000. Symposium: Dry matter intake of lactating dairy  
1034 cattle. *J. Dairy Sci.* 83, 1573-1597.
- 1035 Ingvarsen, K.L., 1994. Models of voluntary food intake in cattle. *Livest. Prod. Sci.* 39,  
1036 19-38.
- 1037 Ingvarsen, K.L., Andersen, H.R., Foldager, J., 1992. Effect of Sex and Pregnancy on  
1038 Feed Intake Capacity of Growing Cattle. *Acta Agr. Scand. A-An.* 42, 40-46.
- 1039 Kitazawa, T., Kaiya, H., 2019. Regulation of gastrointestinal motility by motilin and  
1040 ghrelin in vertebrates. *Front. Endocrinol.* 10, 278.
- 1041 Leão, M.I., 2002. Metodologias de coletas de digestas omasal e abomasal em novilhos  
1042 submetidos a três níveis de ingestão: consumo, digestibilidade e produção  
1043 microbiana (Doctoral dissertation). Federal University of Minas Gerais, Belo  
1044 Horizonte, MG, Brazil.
- 1045 Linden, D., Titgemeyer, E., Olson, K., Anderson, D., 2014. Effects of gestation and  
1046 lactation on forage intake, digestion, and passage rates of primiparous beef  
1047 heifers and multiparous beef cows. *J. Anim. Sci.* 92, 2141-2151.
- 1048 Lopez, S., Hovell, F., Dijkstra, J., France, J., 2003. Effects of volatile fatty acid supply  
1049 on their absorption and on water kinetics in the rumen of sheep sustained by  
1050 intragastric infusions. *J. Anim. Sci.* 81, 2609-2616.

- 1051 Mariz, L.D.S., Amaral, P.M., Valadares Filho, S.C., Santos, S.A., Marcondes, M.I.,  
1052 Prados, L.F., Pacheco, M.V.C., Zanetti, D., Menezes, G.C.C., Faciola, A.P.,  
1053 2018. Dietary protein reduction on microbial protein, amino acids digestibility,  
1054 and body retention in beef cattle. I. Digestibility sites and ruminal synthesis  
1055 estimated by purine bases and <sup>15</sup>N as markers. *J. Anim. Sci.* 96, 2453-2467.
- 1056 Meyer, A.M., Reed, J.J., Vonnahme, K.A., Soto-Navarro, S.A., Reynolds, L.P., Ford,  
1057 S.P., Hess, B.W., Caton, J.S., 2010. Effects of stage of gestation and nutrient  
1058 restriction during early to mid-gestation on maternal and fetal visceral organ  
1059 mass and indices of jejunal growth and vascularity in beef cows. *J. Anim. Sci.*  
1060 88, 2410-2424.
- 1061 Moyo, M., Nsahlai, I.V., 2018. Rate of Passage of Digesta in Ruminants; Are Goats  
1062 Different?, in: Kukovics, S. (Eds.), *Goat science*, IntechOpen. London, UK, pp.  
1063 39-74
- 1064 NASEM, 2016. *Nutrient requirements of beef cattle*. The National Academies Press  
1065 Washington, DC.
- 1066 Okine, E.K., Mathison, G.W., 1991. Reticular contraction attributes and passage of  
1067 digesta from the ruminoreticulum in cattle fed roughage diets. *J. Anim. Sci.* 69,  
1068 2177-2186.
- 1069 Prates, L., Valadares, R., Valadares Filho, S., Detmann, E., Santos, S., Braga, J.,  
1070 Pellizzoni, S., Barbosa, K., 2012. Endogenous fraction and urinary recovery of  
1071 purine derivatives in Nellore and Holstein heifers with abomasal purine infusion.  
1072 *Livest. Sci.* 150, 179-186.
- 1073 Ribeiro, R.C.O., Villela, S.D.J., Valadares Filho, S.C., Santos, S.A., Ribeiro, K.G.,  
1074 Detmann, E., Zanetti, D., Martins, P.G.M.A., 2015. Effects of roughage sources



- 1075 produced in a tropical environment on forage intake, and ruminal and microbial  
1076 parameters. *J. Anim. Sci.* 93, 2363-2374.
- 1077 Robinson, J., 1986. Changes in body composition during pregnancy and lactation. *P.*  
1078 *Nutr. Soc.* 45, 71-80.
- 1079 Robinson, P., Tamminga, S., Van Vuuren, A., 1987. Influence of declining level of feed  
1080 intake and varying the proportion of starch in the concentrate on rumen ingesta  
1081 quantity, composition and kinetics of ingesta turnover in dairy cows. *Livest.*  
1082 *Prod. Sci.* 17, 37-62.
- 1083 Rotta, P., Valadares Filho, S., Gionbelli, T., Costa e Silva, L., Engle, T., Marcondes,  
1084 M., Machado, F., Villadiego, F., Silva, L., 2015. Effects of day of gestation and  
1085 feeding regimen in Holstein x Gyr cows: I. Apparent total-tract digestibility,  
1086 nitrogen balance, and fat deposition. *J. Dairy Sci.* 98, 3197-3210.
- 1087 Santos, S.A., Rotta, P.P., Costa e Silva, L.F., Menezes, A.C.B., Pina, D.S., Valadares  
1088 Filho, S.C, 2016. Protein ruminal degradation of feeds and microbial protein  
1089 synthesis, in: Valadares Filho, S.C. et al. (Eds.), *Nutrient Requirements of Zebu*  
1090 *and Crossbred Cattle - BR-CORTE 3.0*. Suprema, Viçosa, Brazil, pp. 43-84.
- 1091 Scheaffer, A., Caton, J., Bauer, M., Reynolds, L., 2001. Influence of pregnancy on body  
1092 weight, ruminal characteristics, and visceral organ mass in beef heifers. *J. Anim.*  
1093 *Sci.* 79, 2481-2490.
- 1094 Stafford, K., 1991. Ruminoreticular motility in ewes during pregnancy and lactation. *J.*  
1095 *Vet. Sci.* 38, 798-800.
- 1096 Stanley, T., Cochran, R., Vanzant, E., Harmon, D., Corah, L., 1993. Periparturient  
1097 changes in intake, ruminal capacity, and digestive characteristics in beef cows  
1098 consuming alfalfa hay. *J. Anim. Sci.* 71, 788-795.

- 1099 Udén, P., Colucci, P.E., Van Soest, P.J., 1980. Investigation of chromium, cerium and  
1100 cobalt as markers in digesta. Rate of passage studies. *J. Sci. Food Agr.* 31, 625-  
1101 632.
- 1102 Valadares Filho, S.C., Costa e Silva, L.F., Gionbelli, M.P., Rotta, P.P., Marcondes,  
1103 M.I., Chizzotti, M.L., Prados, L.F. 2016. *Nutrient Requirements of Zebu and*  
1104 *Crossbred Cattle - BR-CORTE 3.0*, third ed. Suprema, Viçosa, Brazil.
- 1105 Van Soest, P.v., Robertson, J., Lewis, B., 1991. Methods for dietary fiber, neutral  
1106 detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *J.*  
1107 *Dairy Sci.* 74, 3583-3597.
- 1108 Vanzant, E.S., Cochran, R.C., Johnson, D.E., 1991. Pregnancy and lactation in beef  
1109 heifers grazing tallgrass prairie in the winter: influence on intake, forage  
1110 utilization, and grazing behavior. *J. Anim. Sci.* 69, 3027-3038.
- 1111 Whittet, K.M., Watson, A.K., Erickson, G.E., Klopfenstein, T.J., 2019. Factors affecting  
1112 urinary creatinine in heifers and cows. *Trans. Anim. Sci.* 3, 532-540.

**Table 1**

Ingredients and chemical composition of the corn silage, concentrate, and experimental diet.

Ingredient, g/kg of dry matter	Corn silage	Concentrate	Diet
Corn silage	100.0	-	959.5
Ground corn	-	239.5	9.7
Soybean meal	-	194.3	7.9
Mineral mixture <sup>1</sup>	-	377.5	15.3
Urea + ammonium sulfate <sup>2</sup>	-	188.7	7.6
Chemical composition, g/kg of dry matter			
Dry matter	303.7	938.5	329.5
Organic matter	953.3	605.6	939.2
Crude protein	63.5	647.3	87.2
apNDF	582.9	55.7	561.6
iNDF	209.7	8.5	201.5
Non-fibrous carbohydrates	277.6	233.1	275.8
Ether extract	29.2	13.3	28.6

apNDF = ash- and protein-free neutral detergent fiber, iNDF = indigestible neutral detergent fiber.

<sup>1</sup> Composition: calcium = 100 g/kg; phosphorus = 40 g/kg; sodium = 165 g/kg; sulfur = 6,000 mg/kg; magnesium = 5,000 mg/kg; copper = 680 mg/kg; zinc = 2,580 mg/kg; fluorine = 400 mg/kg; manganese = 750 mg/kg; iron = 350 mg/kg; selenium = 7 mg/kg; iodine = 45 mg/kg; cobalt = 35 mg/kg.

<sup>2</sup> Mixture of urea and ammonium sulfate (9:1 ratio).

**Table 2**

Effect of physiological status (PS) and days of pregnancy (DOP) on body weight (BW) and body condition score (BCS) of beef heifers.

Item	Physiological status		SEM	P-value		
	Non-pregnant	Pregnant		PS	DOP	PSxDOP
Initial BW, kg	466	460	54	0.93	-	-
Initial BCS	6.04	5.93	0.33	0.80	-	-
BW, kg	497	510	10	0.30	<0.01	0.22
BWnp, kg	497	477	10	0.16	<0.01	<0.01
SBW, kg	485	499	10	0.29	<0.01	0.23
SBWnp, kg	485	466	10	0.16	<0.01	<0.01
PREG <sup>1</sup> , kg	-	33.0	3.7	-	<0.01	-
BCS	6.10	5.87	0.30	0.47	0.90	0.65

PSxDOP = interaction between physiological status and days of pregnancy; BWnp = maternal BW (subtracting gravid uterus weight and udder accretion); SBW = shrunk body weight; SBWnp = maternal SBW (subtracting gravid uterus weight and udder accretion); PREG = pregnancy component.

<sup>1</sup> Gravid uterus minus the non-pregnant uterus plus the accretion in udder related to pregnancy.

**Table 3**

Effect of physiological status (PS) and days of pregnancy (DOP) on intake and intake in relation to body weight (BW) of beef heifers.

Item	Physiological status		SEM	P-value		
	Non-pregnant	Pregnant		PS	DOP	PS×DOP
Intake, kg/d						
Dry matter	6.64	6.76	0.45	0.84	<0.01	<0.01
Organic matter	6.33	6.39	0.43	0.92	<0.01	<0.01
Crude protein	0.60	0.61	0.04	0.89	<0.01	0.01
apNDF	3.64	3.68	0.23	0.88	<0.01	0.04
iNDF	1.33	1.35	0.09	0.86	<0.01	0.08
NFC	2.01	2.03	0.14	0.91	<0.01	0.25
Ether extract	0.21	0.21	0.02	0.93	<0.01	0.14
TDN	3.93	3.94	0.22	0.99	0.01	0.06
Intake, g/kg of BW						
Dry matter	13.4	13.5	0.83	0.99	<0.01	<0.01
apNDF	7.29	7.30	0.47	0.99	<0.01	<0.01
iNDF	2.66	2.68	0.18	0.94	<0.01	0.02
TDN	7.94	7.80	0.43	0.80	<0.01	0.05

PS×DOP = interaction between physiological status and days of pregnancy; apNDF = ash- and protein-free neutral detergent fiber; iNDF = indigestible neutral detergent fiber; NFC = non-fibrous carbohydrates; TDN = total digestible nutrient.

**Table 4**

Effect of physiological status (PS) and days of pregnancy (DOP) on ruminal, intestinal, and apparent total-tract digestibility of beef heifers.

Item	Physiological status		SEM	P-value		
	Non-pregnant	Pregnant		PS	DOP	PSxDOP
Ruminal digestibility, g/kg of DM						
Dry matter	405	378	19	0.22	<0.01	0.19
Organic matter	483	464	19	0.09	<0.01	0.19
Crude protein	94.6	71.1	32.0	0.58	<0.01	0.51
apNDF	366	336	19	0.02	<0.01	0.26
NFC	789	802	12	0.37	0.83	0.78
Intestinal digestibility, g/kg of DM reaching the omasum						
Dry matter	271	272	36	0.98	0.77	0.98
Organic matter	244	239	33	0.90	0.58	0.95
Crude protein	519	541	25	0.52	0.18	0.38
apNDF	164	146	17	0.39	0.85	0.37
NFC	558	600	38	0.41	0.33	0.56
Apparent total-tract digestibility, g/kg of dry matter						
Dry matter	554	537	9	0.09	<0.01	0.56
Organic matter	598	576	13	0.11	<0.01	0.67
Crude protein	551	577	7	0.01	<0.01	0.04
apNDF	452	401	8	<0.01	<0.01	0.42
NFC	898	913	12	0.19	0.89	0.35
TDN	600	581	15	0.24	<0.01	0.72

PSxDOP = interaction between physiological status and days of pregnancy; apNDF = ash- and protein-free neutral detergent fiber; NFC = non-fibrous carbohydrates; TDN = total digestible nutrient.

**Table 5**

Effect of physiological status (PS) and days of pregnancy (DOP) on ruminal pool, outflow, rates of ingestion, passage, and digestion, and retention time of beef heifers

Item	Physiological status		SEM	P-value		
	Non-pregnant	Pregnant		PS	DOP	PS×DOP
Ruminal pool, kg of wet matter	34.4	31.8	1.5	0.20	<0.01	0.01
Ruminal dry matter						
Pool, kg	5.25	5.09	0.27	0.66	0.30	<0.01
Outflow, kg/day	3.95	4.48	0.19	0.06	<0.01	0.10
ki, %/h	5.43	6.33	0.17	<0.01	0.21	0.05
kp, %/h	3.20	3.74	0.15	<0.01	<0.01	0.01
kd, %/h	2.25	2.60	0.11	0.05	<0.01	0.34
Retention time, h	31.8	27.4	1.2	<0.01	<0.01	0.23
Ruminal apNDF						
Pool, kg	3.79	3.81	0.21	0.92	<0.01	0.02
Outflow, kg/day	2.32	2.52	0.11	0.21	0.03	<0.01
ki, %/h	3.95	4.45	0.14	0.01	<0.01	0.09
kp, %/h	2.57	2.94	0.15	<0.01	<0.01	0.22
kd, %/h	1.41	1.54	0.07	0.25	0.07	0.04
Retention time, h	40.0	35.6	2.1	<0.01	<0.01	0.74

PS×DOP = interaction between physiological status and days of pregnancy; apNDF =

ash- and protein-free neutral detergent fiber; ki = ingestion rate; kp = passage rate; kd

= digestion rate.

**Table 6**

Effect of physiological status (PS) and days of pregnancy (DOP) on nitrogen (N) balance and microbial crude protein synthesis of beef heifers.

Item	Physiological status		SEM	P-value		
	Non-pregnant	Pregnant		PS	DOP	PSxDOP
Nitrogen intake, g/day	95.5	96.6	5.4	0.88	<0.01	0.10
Fecal nitrogen						
g/day	45.8	44.9	2.6	0.80	0.27	0.21
% of N intake	47.8	46.6	0.8	0.25	0.18	0.57
Urinary nitrogen						
g/day	42.5	41.6	2.1	0.73	0.01	0.38
% of nitrogen intake	46.4	45.6	3.1	0.83	0.35	0.17
Total nitrogen excretion						
g/day	88.5	86.4	4.1	0.70	0.01	0.60
% of nitrogen intake	95.3	92.2	2.8	0.43	0.68	0.21
Nitrogen balance						
g/day	7.47	8.82	2.36	0.66	0.60	0.46
% of N intake	6.82	7.36	2.95	0.89	0.61	0.34
Ruminal outflow nitrogen						
g/day	88.3	88.9	7.5	0.90	<0.01	0.03
% of nitrogen intake	93.5	91.5	3.0	0.61	<0.01	0.55
Ruminal nitrogen balance						
g/day	6.55	9.09	2.70	0.49	<0.01	0.55
% of nitrogen intake	6.49	8.50	2.95	0.61	<0.01	0.55
Efficiency of microbial crude protein synthesis						
g/day	459	461	27	0.95	0.04	0.96
g/kg crude protein intake	797	801	23	0.90	<0.01	0.02
g/kg DOM intake	128	133	5	0.44	<0.01	0.09
g/kg TDN intake	122	125	4	0.65	<0.01	0.09

PSxDOP = interaction between physiological status and days of pregnancy; DOM =

digestible organic matter; TDN = total digestible nutrient.



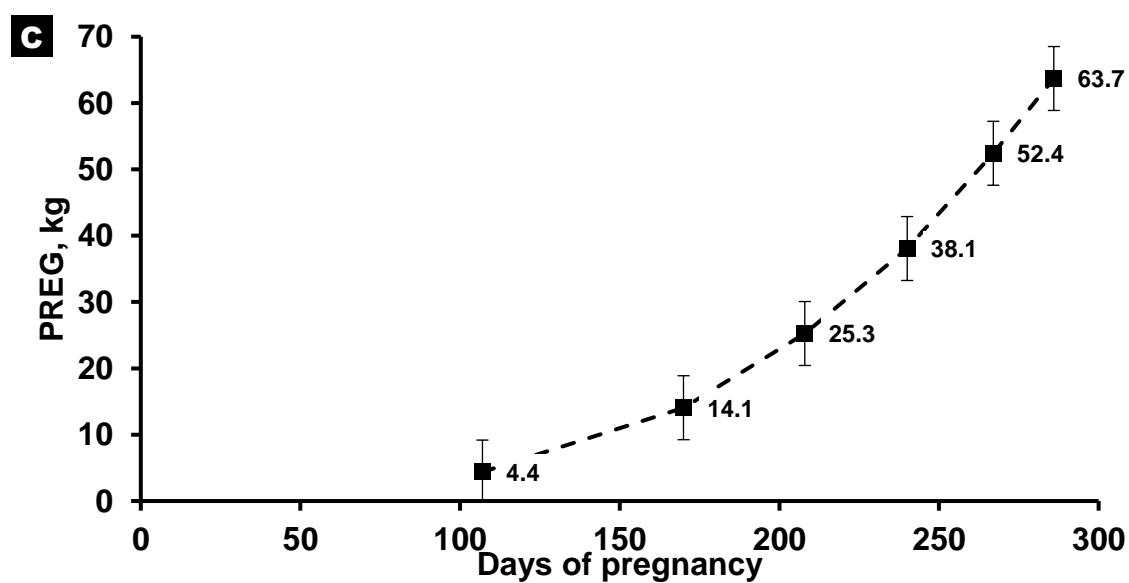
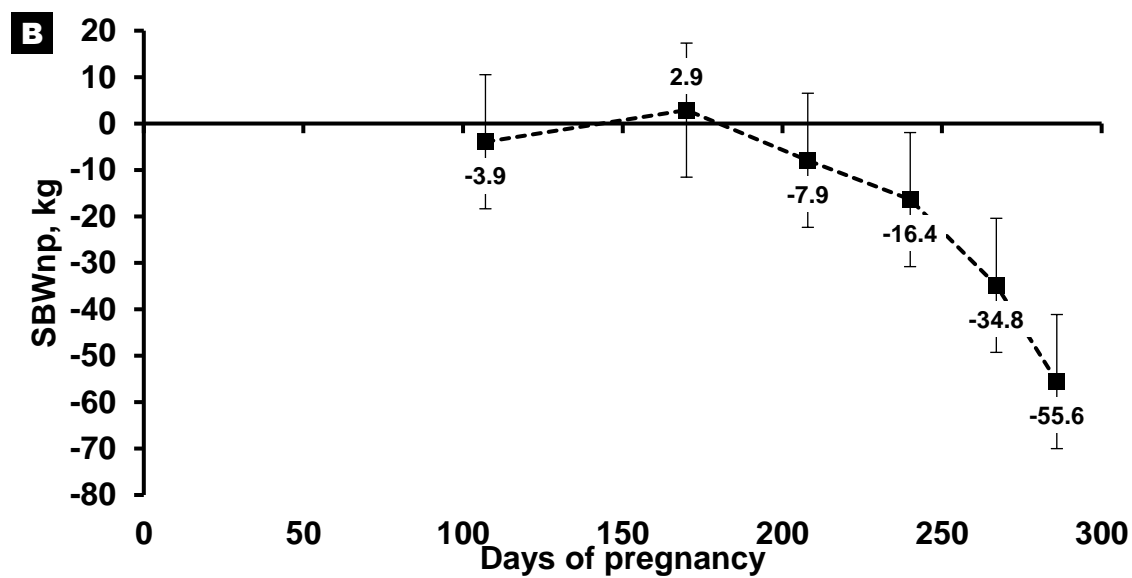
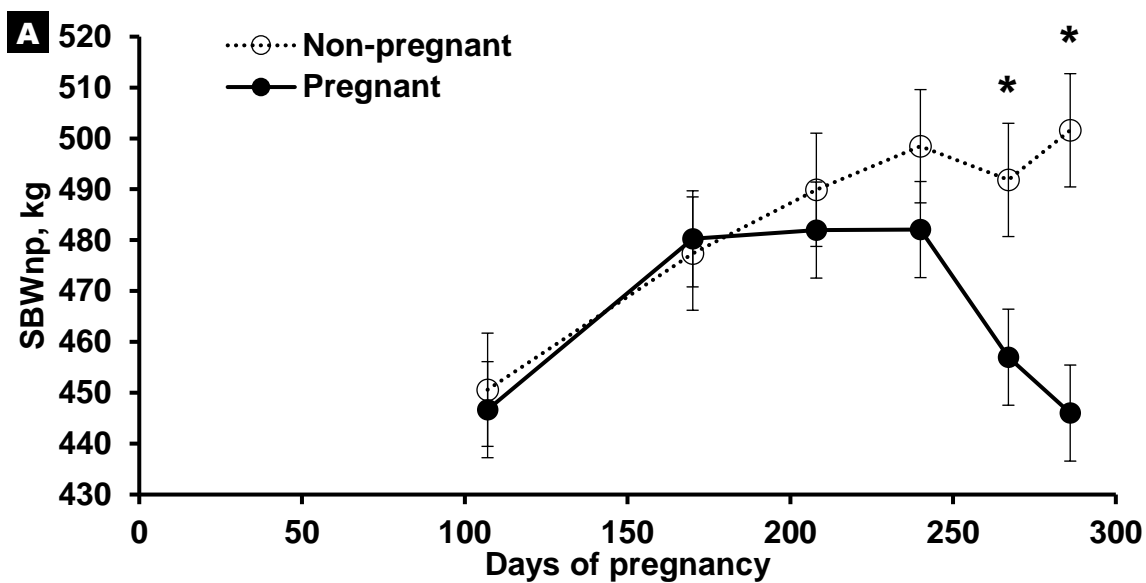
**Table 7**

Effect of physiological status (PS) and days of pregnancy (DOP) on ruminal fermentation parameters of beef heifers.

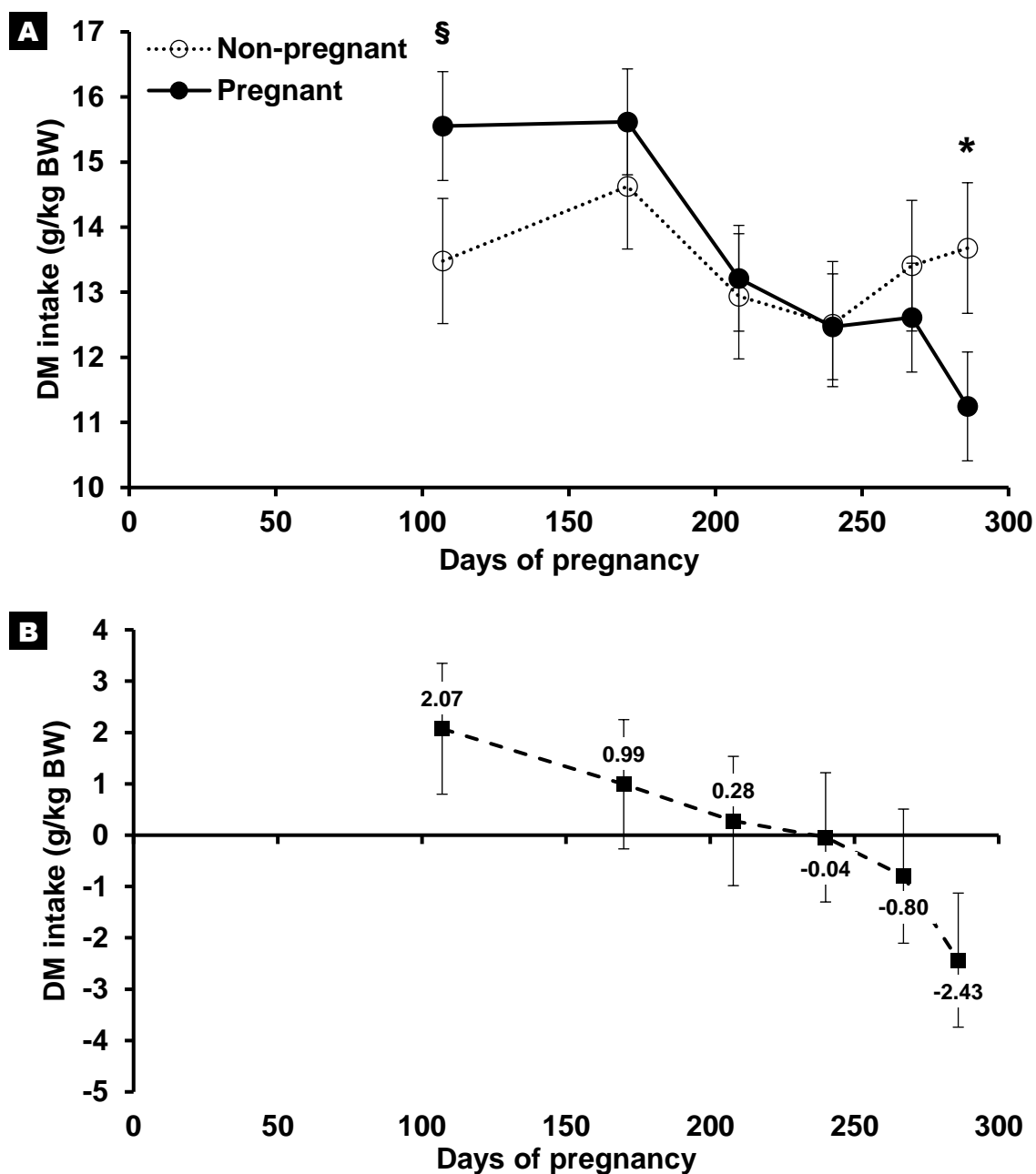
Item	Physiological status		SEM	P-value		
	Non-pregnant	Pregnant		PS	DOP	PS×DOP
Before morning feeding						
pH	7.01	6.99	0.10	0.79	0.08	0.25
N-NH <sub>3</sub> , mg/dL	14.9	15.4	1.0	0.71	0.69	0.34
VFA concentration, mmol/L						
Total VFA	61.4	56.6	11.8	0.66	0.45	0.75
Acetic acid	44.0	41.1	8.5	0.71	0.34	0.72
Propionic acid	14.7	13.2	2.6	0.56	0.97	0.83
Butyric acid	2.21	2.50	0.54	0.55	<0.01	0.97
VFA concentration, % of total of VFA						
Acetic acid	71.5	71.5	1.0	0.96	<0.01	0.19
Propionic acid	24.6	24.0	0.8	0.60	<0.01	0.33
Butyric acid	3.67	4.33	0.34	0.17	<0.01	0.22
A:P ratio	3.03	3.09	0.13	0.77	<0.01	0.16
Four hours after morning feeding						
pH	6.77	6.74	0.13	0.81	0.02	0.78
N-NH <sub>3</sub> , mg/dL	22.9	23.3	1.3	0.76	0.19	0.16
VFA concentration, mmol/L						
Total VFA	60.3	67.7	3.3	0.12	<0.01	0.53
Acetic acid	42.1	46.4	2.4	0.20	<0.01	0.51
Propionic acid	15.3	17.2	0.7	0.06	<0.01	0.14
Butyric acid	2.88	3.06	0.19	0.46	0.14	0.70
VFA concentration, % of total VFA						
Acetic acid	70.0	68.9	1.3	0.52	0.32	0.60
Propionic acid	25.2	25.5	0.4	0.51	0.24	0.16
Butyric acid	4.79	4.51	0.23	0.38	0.80	0.97
A:P ratio	2.76	2.72	0.07	0.60	0.27	0.35

PS×DOP = interaction between physiological status and days of pregnancy; N-NH<sub>3</sub> = ammoniacal nitrogen; VFA = volatile fatty acids; A:P = acetic and propionic acids ratio

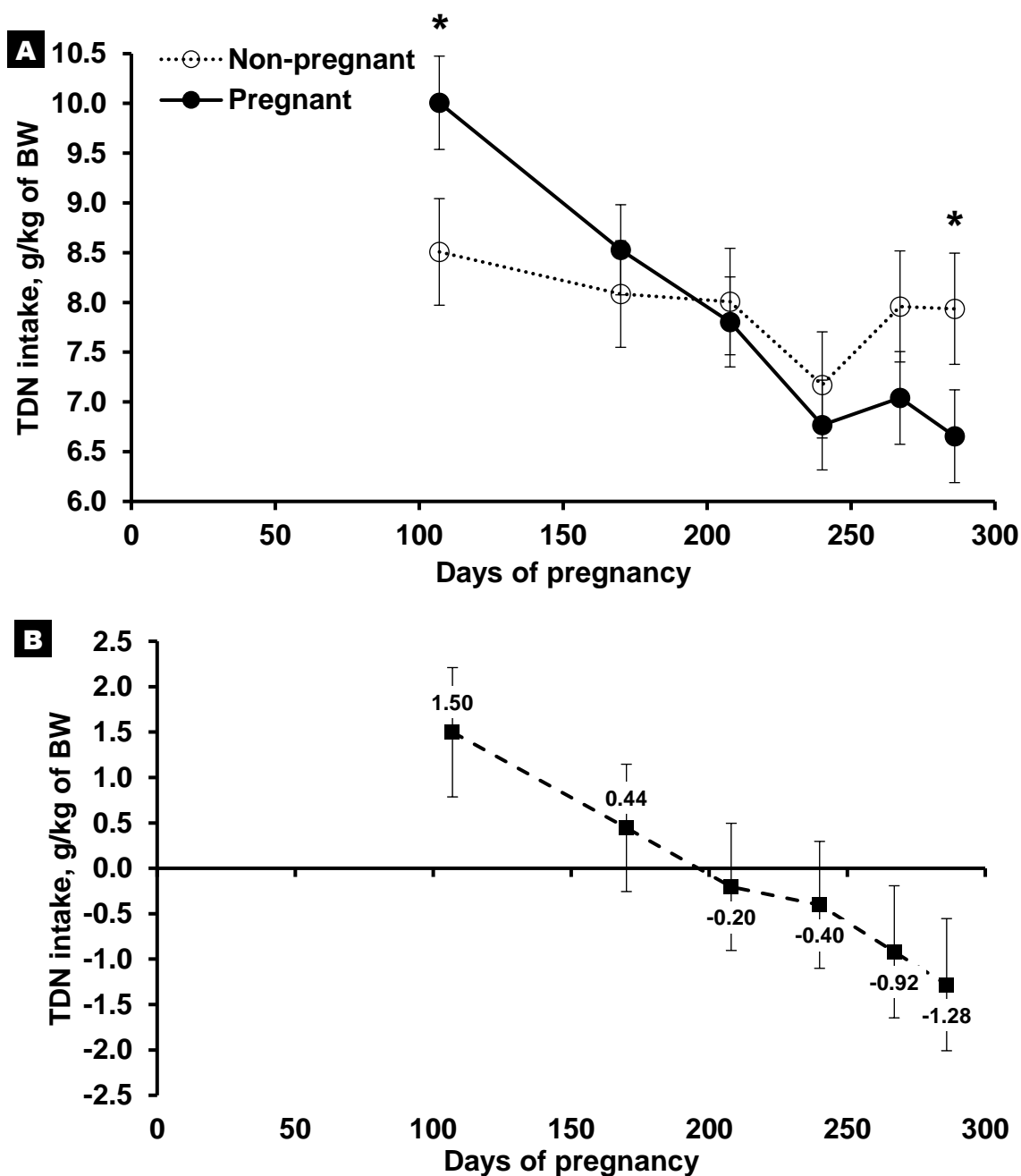
## Figures



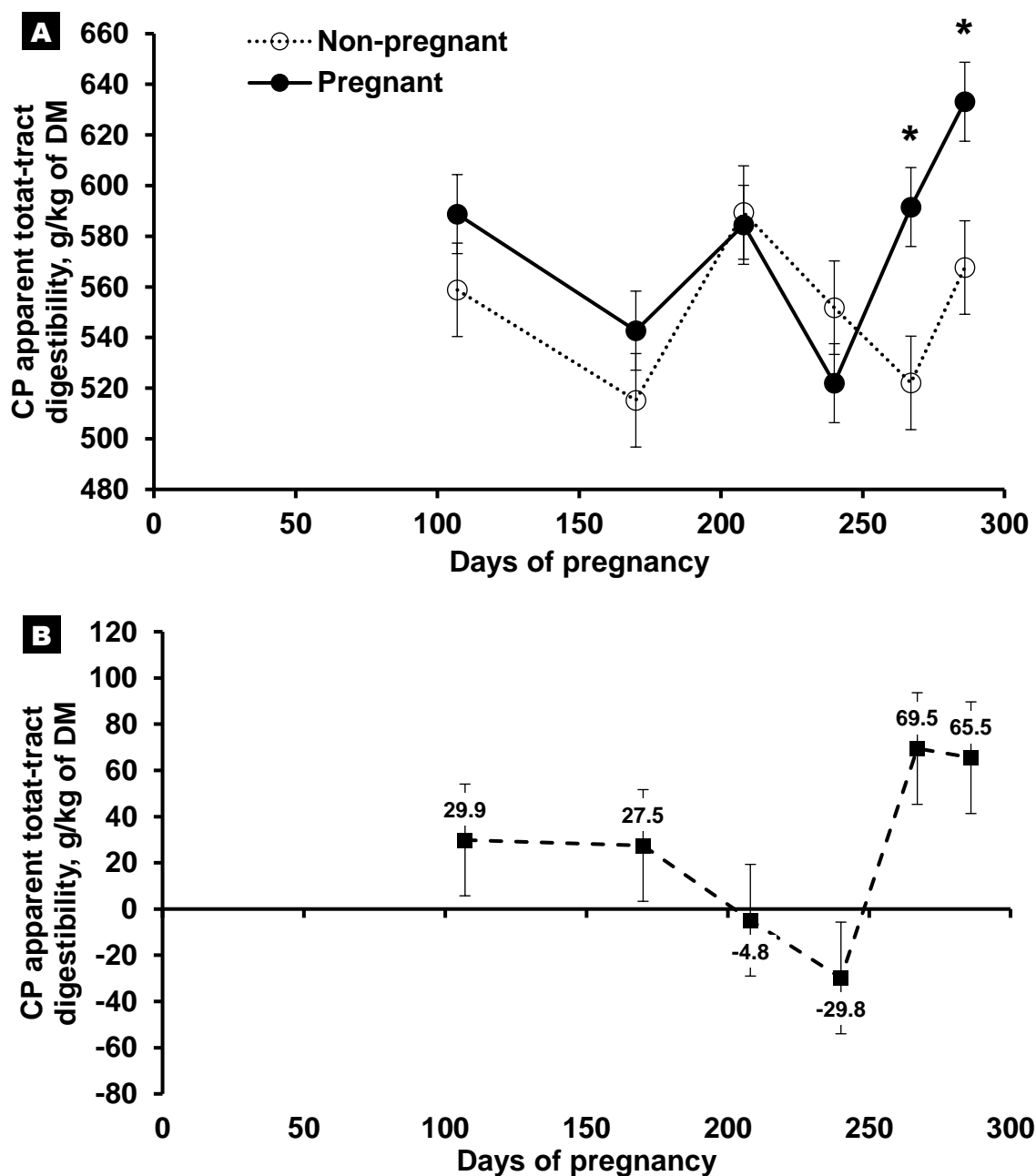
**Fig. 1** Effect of physiological status and days of pregnancy on maternal shrunk body weight of beef heifers. **(A)** Maternal shrunk body weight (SBWnp; kg) of pregnant and non-pregnant beef heifers over days. **(B)** Estimated differences in SBWnp between pregnant and non-pregnant (kg) over days. **(C)** Estimated pregnancy component (kg) of pregnant heifers over days, according to Gionbelli et al. (Gionbelli et al., 2015). \*  $P \leq 0.10$ .



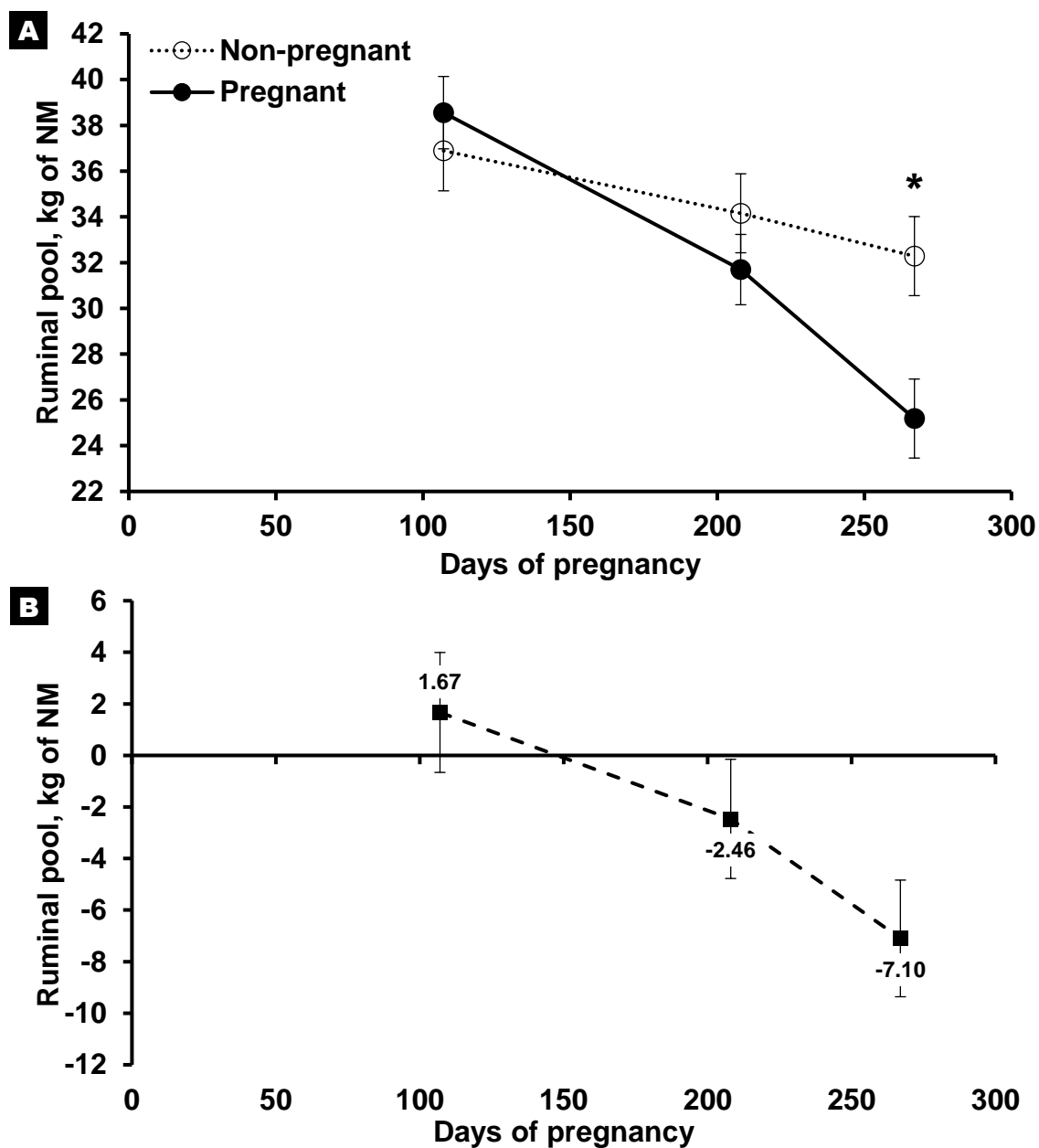
**Fig. 2** Effect of physiological status and days of pregnancy on dry matter intake of beef heifers. **(A)** Dry matter (DM) intake of pregnant and non-pregnant beef heifers (g/kg of BW) over days. **(B)** Estimated differences in DM intake (g/kg of BW) between pregnant and non-pregnant beef heifers over days. §  $P < 0.15$ ; \*  $P \leq 0.10$ .



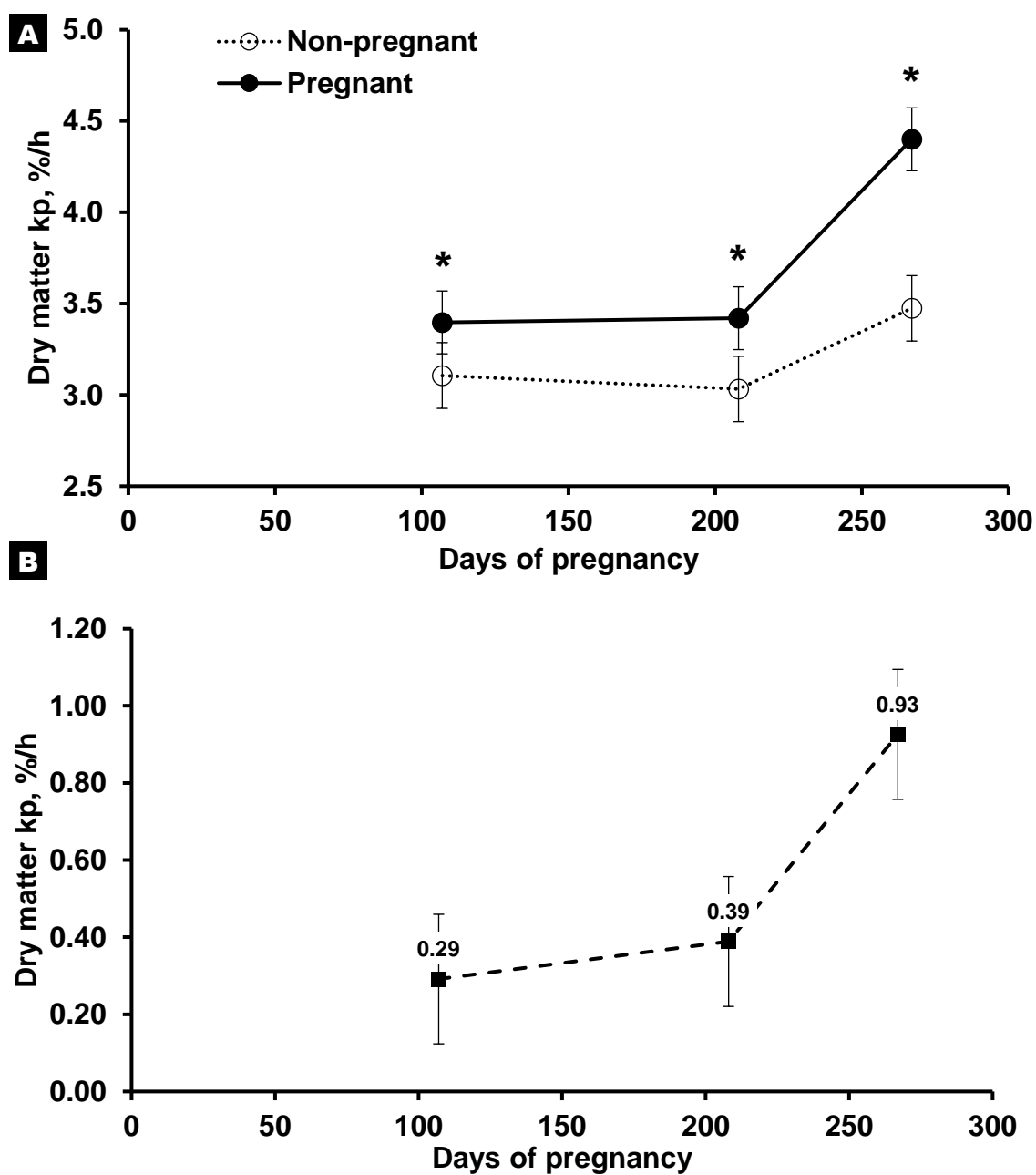
**Fig. 3** Effect of physiological status and days of pregnancy on total digestible nutrients intake of beef heifers. **(A)** Total digestible nutrients (TDN) intake of pregnant and non-pregnant beef heifers (g/kg of BW) over days. **(B)** Estimated differences in TDN intake (g/kg of BW) between pregnant and non-pregnant beef heifers over days. \*  $P \leq 0.10$ .



**Fig. 4** Effect of physiological status and days of pregnancy on crude protein apparent total-tract digestibility of beef heifers. **(A)** Crude protein apparent total-tract digestibility of pregnant and non-pregnant beef heifers (g/kg of DM) over days. **(B)** Estimated differences in crude protein apparent total-tract digestibility (g/kg of DM) between pregnant and non-pregnant beef heifers over days. \*  $P \leq 0.10$ .

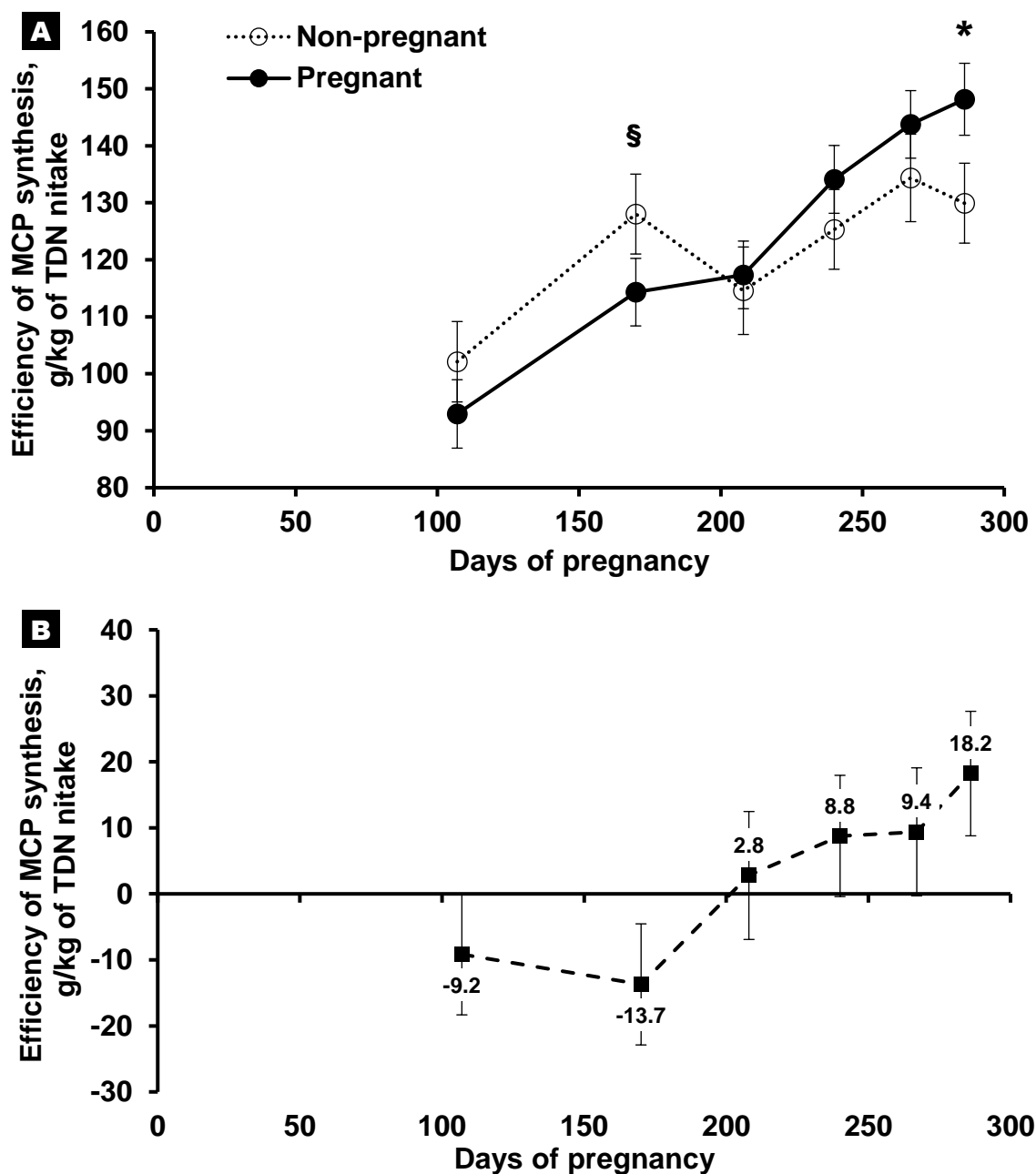


**Fig. 5** Effect of physiological status and days of pregnancy on the ruminal pool of wet matter of beef heifers. **(A)** Ruminal pool of wet matter (NM) of pregnant and non-pregnant beef heifers (kg of NM) over days. **(B)** Estimated differences in ruminal pool of wet matter (kg) between pregnant and non-pregnant beef heifers over days. \*  $P \leq 0.10$ .



**Fig. 6** Effect of physiological status and days of pregnancy on dry matter passage rate of beef heifers. **(A)** Dry matter (DM) passage rate (kp) of pregnant and non-pregnant beef heifers (%/h) over days. **(B)** Estimated differences in DM kp (%/h) between pregnant and non-pregnant beef heifers over days. \*  $P \leq 0.10$ .





**Fig. 7** Effect of physiological status and days of pregnancy on the efficiency of microbial crude protein synthesis of beef heifers. **(A)** Efficiency of microbial crude protein (MCP) synthesis of pregnant and non-pregnant beef heifers (g/kg of TDN intake) over days. **(B)** Estimated differences in efficiency of MCP synthesis (g/kg of TDN intake) between pregnant and non-pregnant beef heifers over days. §  $P < 0.15$ ; \*  $P \leq 0.10$ .

1156 **ARTICLE 2 - The course of pregnancy changes general metabolism and affects**  
1157 **ruminal epithelium activity pattern in zebu beef heifers**

1158

1159 Article formatted according to Livestock Science guidelines

1160

1161 **The course of pregnancy changes general metabolism and affects ruminal**  
1162 **epithelium activity pattern in zebu beef heifers**

1163

1164 **Abstract**

1165 The present study aimed to investigate metabolic and physiological changes induced  
1166 by pregnancy, as well as the expression of key genes linked to the absorptive and  
1167 proliferative activity of the ruminal epithelium towards the advance of the gestational  
1168 period in zebu beef heifers. Twelve ruminally cannulated Zebu beef heifers were  
1169 randomly assigned into two experimental treatments: a pregnant group (n = 7) and a  
1170 control group (non-pregnant; n = 5). All heifers received the same diet throughout the  
1171 experiment. Respiratory and heart rates and plasma glucose levels were assessed just  
1172 before and four hours after morning feeding at 110, 171, 206, 242, 266, and 286 days  
1173 of pregnancy (DOP). Blood samples were collected for serum non-esterified fatty acids  
1174 (NEFA), beta-hydroxybutyrate (BHB), and urea analysis prior to the morning feedings  
1175 at 110, 206, 266, and 286 DOP. At 215 and 272 DOP the ruminal epithelium was  
1176 biopsied to evaluate the mRNA expression of key genes involved in remodeling,  
1177 inflammation, and transport. The respiratory rate was similar ( $P \geq 0.79$ ) between  
1178 groups over days. However, the heart rate increased in late-pregnant heifers compared  
1179 to controls. Blood concentrations of NEFA ( $P = 0.11$ ) and BHB ( $P = 0.67$ ) did not vary  
1180 over the gestation period in pregnant heifers. Glucose levels before morning feeding

1181 were similar during all collection periods with an exception at 286 days (DOP ×  
1182 physiological status interaction;  $P = 0.05$ ) when glucose was lower in pregnant (83  
1183 mg/dL) compared to non-pregnant (107 mg/dL) heifers. The mRNA expression of  
1184 genes related to cellular remodeling (*PCNA* and *CASP3*), inflammation (*KLK9* and  
1185 *KLK10*), and transport of volatile fatty acids (*SLC16A1* and *SLC16A3*),  $H^+$  (*SLC9A1*),  
1186  $HCO_3^-$  (*SLC26A3* and *SLC26A6*), and glucose (*SLC5A1*) in the ruminal epithelium  
1187 were downregulated at late gestation. These results suggest that the ruminal  
1188 epithelium saves energy at late pregnancy to benefit fetal development. In addition,  
1189 the increase in heart rate coupled with tissue mobilization can be considered  
1190 homeorhetic mechanisms that help meet fetal nutrient requirements.

1191

1192 **Keywords:** gene expression, heart rate, homeorhesis, rumen epithelium, Zebu cattle

1193

## 1194 1. Introduction

1195 Regulation of nutrient partitioning during pregnancy involves homeorhetic  
1196 controls arising from the conceptus (Bauman and Currie, 1980). It is suggested that  
1197 late-gestation pregnant cows may be able to reduce maintenance energy costs to  
1198 support the energetic demands of the conceptus (Freetly et al., 2008). Even though  
1199 gestation is a physiologic state (PS) distinguished by increased nutritional  
1200 requirements, a decrease in dry matter (DM) intake is usually observed as gestation  
1201 advances (Gionbelli et al., 2016).

1202 Furthermore, as pregnancy advances and the concept grow, the enlarged  
1203 uterus compresses maternal organs such as the rumen (Forbes, 1968). The reduction  
1204 in ruminal capacity can induce a decrease in feed intake (Hanks et al., 1993; Stanley  
1205 et al., 1993; Scheaffer et al., 2001). This physical limitation can often be compensated

1206 by an increase in the feed passage rate, hence allowing maintenance of the intake  
1207 level. Still, the faster feed passes through the digestive tract; the smaller tends to be  
1208 its digestibility (Ribeiro et al., 2015). As passage rate and digestibility change, other  
1209 parameters of ruminal fermentation may also be modified (Hare et al., 2019).  
1210 Production and absorption of volatile fatty acids (VFA), nitrogen compounds usage,  
1211 microbial protein synthesis, and balance of ruminal pH are all subject to change as  
1212 gestation progresses.

1213 The ruminal epithelium has an essential role in the absorption VFA as well as  
1214 the regulation of luminal pH (Aschenbach et al., 2011). In dairy cattle, ruminal  
1215 epithelium transcriptome in the transition period has been a target in several studies  
1216 (Minuti et al., 2015; Steele et al., 2015; Bach et al., 2018). However, both the type and  
1217 amount of feed consumed by lactating and dry cows differ drastically in dairy cows,  
1218 while in beef cattle, the changes are slighter. Thus, information about the pattern of  
1219 activity of the ruminal epithelium during the gestation of beef cattle may lead to an  
1220 increased understanding of cellular mechanisms involved in energetic partitioning.

1221 Therefore, the present study aimed to investigate possible metabolic and  
1222 physiological changes induced by pregnancy, as well as the mRNA expression of key  
1223 genes involved in the absorptive and proliferative activity of the ruminal epithelium  
1224 towards the advance of the gestational period in zebu beef heifers. It was hypothesized  
1225 that gestation advancement promotes homeorhetic changes to reduce maternal  
1226 maintenance energy expenditure.

1227

## 1228 **2. Material and Methods**

1229 This study was conducted at the Department of Animal Sciences facilities of the  
1230 Federal University of Lavras (UFLA), Lavras, Minas Gerais, Brazil. All the procedures

1231 involving the use of animals in this study followed the guidelines established by the  
1232 UFLA Ethics Committee on Animal Use (protocol number: 048/16).

1233

### 1234 2.1. *Animals, housing, and feeding*

1235 Animal handling procedures have been previously reported (Moreira et al.,  
1236 2020). Briefly, 12 ruminally cannulated Zebu heifers (BW = 417 ± 95.6 kg) were  
1237 randomly assigned into two groups, a pregnant group (n = 7) and a control group (non-  
1238 pregnant; n = 5). Heifers assigned to the pregnant group were submitted to an  
1239 ovulation synchronization protocol and the day zero of gestation was considered the  
1240 day of artificial insemination. On day 85 of gestation of the pregnant group, all heifers  
1241 were housed in individual pens (80 m<sup>2</sup>). After penning, heifers started the experimental  
1242 adaptation phase. The feed intake control started on the 95<sup>th</sup> day of gestation and  
1243 ended at calving.

1244 The experimental diet [DM = 32.95%; crude protein (CP) = 8.72%, and neutral  
1245 detergent fiber (NDF) = 56.16%] was formulated according to the Nutrient  
1246 Requirements of Zebu and Crossbred Cattle - BR-CORTE (Valadares Filho et al.,  
1247 2016), to allow *ad libitum* intake without large accumulation of body reserves and  
1248 adequate maintenance of gestation. The experimental diet (DM basis) was based on  
1249 95.95% of medium quality corn silage (DM = 30.37%; CP = 6.35%, and NDF =  
1250 58.29%), and 4.05% of concentrate supplement (DM = 93.85%; CP = 64.73%, and  
1251 NDF = 5.57%) prepared from ground corn (0.97%), soybean meal (0.79%), urea plus  
1252 ammonium sulfate (0.76%), and macro and micro mineral mixture (1.53%). All heifers  
1253 received the same diet throughout the experiment. Animals were fed twice a day at  
1254 0800 and 1400 h.

1255

1256 2.2. *Data collection and analysis*

1257 A timeline of sampling days was followed during the experiment (Fig. 1).

1258

1259 2.2.1. *Respiratory and heart rates evaluation*

1260 Respiratory and heart rates were assessed just before and four hours after  
1261 morning feedings at 110, 171, 206, 242, 266, and 286 days of pregnancy (DOP).

1262 Respiratory rates (RR) were monitored for 1 min by counting flank movements (Milan  
1263 et al., 2016). Heart rates (HR) were determined by counting heartbeats with a  
1264 stethoscope placed on the left chest wall over the cardiac area for 1 min (Ghizzi et al.,  
1265 2018).

1266

1267 2.2.2. *Blood metabolite levels*

1268 Blood samples were collected via coccygeal venipuncture in 4 mL sodium  
1269 fluoride tubes (code: 50205, Labor Import, Osasco, Brazil) for plasma glucose analysis.

1270 For non-esterified fatty acid (NEFA), beta-hydroxybutyrate (BHB), and urea analysis a  
1271 10 mL spray-coated silica tube (code: 50208, Labor Import, Osasco, Brazil) was used.

1272 For NEFA, BHB, and urea analysis, blood samples were obtained just before morning  
1273 feeding at 110, 206, 266, and 286 DOP. While for glucose analysis, samples were  
1274 collected at 110, 171, 206, 242, 266, and 286 DOP just before and four hours after  
1275 morning feedings. Blood was immediately centrifuged at  $2700 \times g$  for 20 min at  $4^{\circ}\text{C}$ .  
1276 Serum/plasma was removed by pipette and frozen at  $-20^{\circ}\text{C}$  until further analyses.

1277 Plasma glucose concentrations were measured according to an enzymatic  
1278 system by a commercial kit (GLICOSE Liquiform, Ref.: 133, Labtest Diagnóstica S.A.,  
1279 Lagoa Santa, Brazil). The NEFA concentrations were measured by Enzyme-Linked  
1280 Immunosorbent Assay (ELISA) according to the recommendations of a commercial kit

1281 (Bovine Non-esterified Fatty Acid ELISA Kit, Bioassay Technology Laboratory,  
1282 Shanghai, China). Serum BHB levels were assayed using a kinetic enzymatic method  
1283 (Ranbut kit no. RB 1007, Randox Laboratory, Antrim, UK). Serum urea was determined  
1284 using an enzymatic system by a commercial kit (Ureia 500, Doles Reagentes e  
1285 Equipamentos para Laboratórios Ltda, Goiânia, Brazil).

1286

### 1287 *2.2.3. Gene expression in ruminal papillae*

1288 At 215 and 272 DOP, total rumen evacuation was performed one hour before  
1289 morning feeding for ruminal papillae collection. The rumen papillae were collected at  
1290 the most ventral site of the ventral rumen sac using curved surgical scissors. Papillae  
1291 were immediately washed with a 0.9% NaCl solution, stored in 2 mL cryotubes, and  
1292 snap-frozen in liquid nitrogen. Papillae samples were stored at  $-80^{\circ}\text{C}$  until RNA  
1293 extraction and mRNA expression analysis.

1294 The design of primers for target and housekeeping genes was performed using  
1295 sequences that are registered and published in the GenBank public data bank, a  
1296 National Center for Biotechnology Information (NCBI) platform (Table 1). Primers were  
1297 designed using OligoPerfect Designer software (Invitrogen, Karlsruhe, Germany) and  
1298 synthesized (Invitrogen, Carlsbad, CA, USA). Total RNA was extracted from papillae  
1299 samples using QIAzol (QIAGEN, Valencia, CA, USA) and treated with DNA-free  
1300 DNase (Ambion, Austin, TX, USA) according to the manufacturer's instructions. To  
1301 analyze the 28S and 18S rRNA bands, the total RNA was electrophoresed in a 1.0%  
1302 (m/v) agarose gel, stained with GelRed nucleic acid gel stain (Biotium, Hayward, CA,  
1303 USA) and visualized with an E-gel Imager Camera Hood (Life Technologies, Neve  
1304 Yamin, Israel). The RNA quantity (ng/ $\mu\text{L}$ ) and purity (260/280 and 260/230) were  
1305 assessed using a spectrophotometer (DeNovix DS-11 Spectrophotometer, USA) at

1306 260 nm. Complementary DNA (cDNA) synthesis was performed using the  
1307 HighCapacity cDNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA,  
1308 USA) according to the manufacturer's instructions, and samples were stored at -20  
1309 °C.

1310 Reverse transcription qPCR (RT-qPCR) was performed on an Eppendorf  
1311 Realplex system (Eppendorf, Hamburg, Germany) with a SYBR Green detection  
1312 system (Applied Biosystems, Foster City, CA, USA). The RT-qPCR reactions were  
1313 performed as the following protocol: 50°C for 2 min, followed by 95°C for 10 min,  
1314 followed by 40 cycles of 95°C for 15 s and 60°C for 1 min. The RT-qPCR analyses of  
1315 each studied gene were performed using cDNA from biological replicates, with two  
1316 technical replicates per biological replicate. The  $\beta$ -actin (*ACTB*; NM\_173979) and  
1317 prefoldin-like chaperone (*UXT*; NM\_001037471) were used as the housekeeping  
1318 genes (Die et al., 2017). Relative expression levels were calculated according to the  
1319 method described by Pfaffl (2001).

1320

### 1321 2.3. *Statistical Analyses*

1322 Data were analyzed through the mixed models methodology (procedure MIXED  
1323 of SAS 9.2, SAS Inst. Inc., Cary, NC), considering PS (pregnant or non-pregnant) and  
1324 DOP as fixed effects and animal as the random effect. When appropriate, initial BW  
1325 was included as a covariate in the model. Once repeated measurements were taken  
1326 within the same animal (for DOP), the subject animal nested treatment was included  
1327 as the error term in the repeated measurement statement. For every DOP, the PS  
1328 effect on the measured variable was estimated using the “estimate statement” of SAS.  
1329 Prior to the final analyses, extreme data were removed when Studentized residuals  
1330 were not within  $\pm 3$  standard deviations, and normality (P-value > 0.10) was assessed



1331 using Shapiro-Wilk's test. As expected, the gene expression data was not normal and  
1332 it was transformed using the RANK procedure of SAS 9.2. The value of 0.10 was  
1333 adopted as a critical level of probability for the occurrence of Type I error.

1334

### 1335 **3. Results and Discussion**

#### 1336 *3.1. Respiratory and heart rates*

1337       Regardless of evaluation time, pregnant and non-pregnant heifers had similar  
1338 RR through gestation (Table 2). However, a DOP effect ( $P = 0.01$ ) was detected four  
1339 hours after feeding. Respiratory rate was reduced over time in pregnant and non-  
1340 pregnant heifers. Variations in RR are mainly linked to environmental fluctuations  
1341 (Hansen, 2004). However, some studies have shown that zebu animals maintain a  
1342 stable respiratory rate regardless of environmental temperatures and reinforce their  
1343 adaptive potential (Melo Costa et al., 2018; Lima et al., 2020). Additionally, although  
1344 previous data have shown that visceral organ mass can be influenced by both  
1345 pregnancy and nutrient intake in pregnant cows (Meyer et al., 2010; Wood et al., 2013;  
1346 Rotta et al., 2015), no differences were found in the lungs or diaphragm weights.

1347       Before morning feedings, a DOP  $\times$  PS interaction effect ( $P = 0.08$ ) was observed  
1348 on HR (Fig. 2). Heart rate was similar until 206 DOP but was greater at late gestation  
1349 (242, 266, and 286 DOP) in pregnant than in non-pregnant heifers. Similarly, four hours  
1350 after morning feedings, HR was greater ( $P < 0.01$ ) in pregnant than in non-pregnant  
1351 heifers. Also, HR increased ( $P < 0.01$ ) through gestation in pregnant, but not in non-  
1352 pregnant heifers (Fig. 2). This increase can be justified by the high demand for nutrients  
1353 and oxygen in maternal tissues and, mainly, uteroplacental (Reynolds et al., 1986).  
1354 Likewise, HR of pregnant heifers gradually increased over time [ $83 \pm 3$  at 14 weeks  
1355 and  $97 \pm 4$  beats/min at 1 week before calving (Trenk et al., 2015)]. According to Brosh

1356 et al. (2002), HR allows estimating energy expenditure in cattle, and the increase  
1357 toward the end of gestation reflects increasing energy demands of the growing fetus.

1358

### 1359 3.2. *Metabolic profile*

1360 In late gestation, conceptus glucose requirements increase by approximately  
1361 50% (Bell, 1995). Despite the importance of glucose for fetal metabolism across  
1362 mammalian species, ruminants exhibit difficulty in increasing its circulatory levels, thus  
1363 making the use of amino acids as precursors for gluconeogenesis as the most  
1364 straightforward path to meet the high uterine glucose demand (McNeill et al., 1997).  
1365 For these reasons, the mobilization of body tissues is compulsory, and consequently,  
1366 the metabolic profile during pregnancy is altered (Linden et al., 2014; Lopes et al.,  
1367 2020). Weight and body condition results are presented in detail elsewhere (Moreira  
1368 et al., 2020). Briefly, despite of no change in body condition score, maternal body  
1369 weight and shrunk body weight (subtracting gravid uterus weight and udder accretion)  
1370 were lower in pregnant compared with non-pregnant heifers at late gestation, indicating  
1371 tissue mobilization at this stage.

1372 Before morning feeding, glucose levels were similar during all collection periods  
1373 with an exception at 286 DOP (DOP × PS interaction;  $P = 0.05$ ) when glucose was  
1374 lower in pregnant than non-pregnant heifers (Table 3 and Fig. 3). Four hours after  
1375 feeding, no differences were found between PS and DOP. A tendency ( $P = 0.11$ ) for a  
1376 PS × DOP interaction effect was detected on NEFA levels. At 286 DOP, NEFA  
1377 concentrations were lower in non-pregnant heifers, whilst similar throughout gestation  
1378 in pregnant heifers. Serum BHB concentration was not affected by PS or DOP. Urea  
1379 concentration (DOP × PS interaction;  $P = 0.03$ ) tended to be lower at 206 DOP (24.3

1380 vs. 21.0 mg/dL) and were greater at 266 DOP (20.4 vs.15.7 mg/dL) in pregnant  
1381 compared to non-pregnant heifers, respectively.

1382         When increased feed intake is not possible, changes in the production and  
1383 utilization of glucose by tissues can be observed (Wood et al., 2013). In the present  
1384 study, feed intake decreased as gestation length increased (Moreira et al., 2020).  
1385 Therefore, high fetal demand for glucose, coupled with low nutrient intake, could be  
1386 the causes of the lower circulating glucose concentration observed at 286 DOP.

1387         Non-esterified fatty acid and BHB are typical markers of fat tissue mobilization  
1388 (Wood et al., 2013). However, since NEFA and BHB concentration, as well as the body  
1389 condition score, did not change during pregnancy, it is suggested that the tissue  
1390 mobilization observed in this study was a consequence of muscle tissue catabolism.  
1391 The mechanisms regulating protein mobilization in late gestation of cows are still not  
1392 clear. Recently, Lopes et al. (2020) found that cows not supplemented during  
1393 pregnancy tended to have greater total circulating amino acids and concentrations of  
1394 circulating glycolytic amino acids than supplemented cows. The authors suggested  
1395 that non-supplemented cows alter their metabolism to meet increasing fetal nutrient  
1396 demands, increasing amino acid mobilization from skeletal muscle tissue for  
1397 gluconeogenesis.

1398

### 1399 3.3. *Gene expression in ruminal papillae*

1400         Pregnant heifers had lower proliferating cell nuclear antigen (*PCNA*) mRNA  
1401 expression than non-pregnant. The protein encoded by this gene is essential for DNA  
1402 replication and can be used as an indicator of cell proliferation. Similarly, cell  
1403 proliferation in the jejunum decreased due to pregnancy in cows (Scheaffer et al.,  
1404 2003), suggesting that the visceral tissue use less energy during pregnancy. On the

1405 other hand, the abundance of proteins encoded by *PCNA* was not affected by  
1406 pregnancy in the rumen papillae (Wood et al., 2013). With a reduced remodeling of the  
1407 ruminal epithelium, it is expected that cell apoptosis will be also reduced (Bach et al.,  
1408 2019). In fact, in the present study, a downregulation in the expression of caspase 3  
1409 (*CASP3*) was verified in pregnant heifers at 272 DOP. The *CASP3* encodes a protease  
1410 that plays a central role in cell apoptosis. Consequently, the decreased cell proliferation  
1411 (based on *PCNA*) and apoptosis (based on *CASP3*) could be indicative of reduced  
1412 rumen epithelium activity at late gestation.

1413         Recent studies have shown that the kallikrein-related peptidases (*KLK*) genes  
1414 are present in the bovine ruminal epithelium and they are related to feed intake, weight  
1415 gain, feed efficiency, and VFA ruminal concentration (Baldwin et al., 2012; Veerkamp  
1416 et al., 2012; Kern et al., 2016a; Kern et al., 2016b). The *KLK* are serine proteases that  
1417 are involved in epidermal processes such as tumor development, regulation of  
1418 inflammation, desquamation of skin cells, and wound healing (Kantyka et al., 2011). It  
1419 is known that the energetic cost of an immune response is high for the animal (Kvidera  
1420 et al., 2017; Reynolds et al., 2017). Thus, a reduction in immune and inflammatory  
1421 responses may allow more energy for maternal and fetal growth. In the study of Kern  
1422 et al. (2016a), the kallikrein-related peptidase 10 (*KLK10*) was found to be upregulated  
1423 in steers with low feed efficiency compared to more efficient animals. In the current  
1424 study, pregnant heifers had lower expression of kallikrein-related peptidase 9 (*KLK9*)  
1425 and *KLK10* at 272 DOP compared to non-pregnant heifers. These results strengthen  
1426 the concept that the ruminal epithelium enters into an energy-saving state at late  
1427 gestation, steadily providing energy for the maintenance of pregnancy.

1428         Ruminal epithelial proliferation has been positively associated with the  
1429 increased surface area for nutrient absorption (Baldwin, 1999). Additionally, an

1430 increase in ruminal epithelium inflammation could reduce the absorption of nutrients  
1431 due to papillae swelling (Kern et al., 2016b). Therefore, the current results suggest that  
1432 pregnant heifers have less surface area for absorption. However, reduced papillae  
1433 swelling can compensate for this diminished absorptive capacity of the ruminal  
1434 epithelium. It is noteworthy that these considerations are based only on gene  
1435 expression since papillae morphology was not assessed.

1436 The mRNA expressions of monocarboxylic acid transporter 1  
1437 (*SLC16A1/MCT1*), sodium–hydrogen antiporter 1 (*SLC9A1/NHE1*), down-regulated in  
1438 adenoma (*SLC26A3/DRA*), putative anion transporter 1 (*SLC26A6/PAT1*), and  
1439 sodium/glucose cotransporter 1 (*SLC5A1/SLGT1*) were affected by a PS × DOP  
1440 interaction ( $P \leq 0.09$ ). The mRNA expression of these genes at 272 DOP were  
1441 decreased in pregnant compared to non-pregnant heifers. Also, the monocarboxylic  
1442 acid transporter 4 (*SLC16A3/MCT4*) mRNA expression was lower in pregnant than in  
1443 non-pregnant heifers. These genes are well known as markers for the regulation of  
1444 VFA,  $H^+$ ,  $HCO_3^-$ , and glucose epithelial transport processes (Connor et al., 2010). The  
1445 MCT4 is responsible for VFA transport from the lumen into epithelial cells. In contrast,  
1446 MCT1 is accountable for transporting VFA from the epithelium into the bloodstream  
1447 (Connor et al., 2010). The mechanism controlling the expression of these genes is still  
1448 unclear. It is suggested that ruminal VFA concentration, especially butyrate, and pH  
1449 can regulate MCT expression (Laarman et al., 2013). However, no difference was  
1450 found in the ruminal VFA profile and pH between the animals in this study (Moreira et  
1451 al., 2020). These results suggest that rumen absorption of VFA by late-pregnant  
1452 heifers is reduced, and other metabolic factors besides rumen fermentation products  
1453 control gene expression in the rumen wall.

1454           Ruminal VFA absorption increases H<sup>+</sup> concentrations in the cytosol (Laarman  
1455 et al., 2016). Thus, the main function of NHE1 is recycling H<sup>+</sup> back to the ruminal lumen  
1456 and importing Na<sup>+</sup> (Connor et al., 2010). On the other hand, the major role of DRA and  
1457 PAT1 in the rumen epithelium is to transport HCO<sub>3</sub><sup>-</sup> from inside the cell to the lumen  
1458 (Connor et al., 2010). The downregulation of these genes suggests that the ruminal  
1459 epithelium reduces the absorption of VFA in late-pregnant heifers. Zhang et al. (2013)  
1460 observed that short-term feed restriction, as occurring in late gestation, has a negative  
1461 effect on acetate absorption. The authors speculated that this acetate absorption  
1462 impairment is a mechanism to reduce the energy required by the ruminal epithelium  
1463 for synthesis of transport proteins when the energy supply is low.

1464

#### 1465 **4. Conclusion**

1466           The heart rate increased throughout gestation in heifers. Also, based on the  
1467 circulating metabolite profiles and body weight loss during the gestational period, it can  
1468 be concluded that pregnant heifers mobilized tissues, especially skeletal muscle.  
1469 Lastly, the current results indicate that the ruminal epithelium of late-pregnant cows  
1470 converts into an energy-saving state, reducing its remodeling and absorptive capacity.  
1471 The current results increase the understanding of the homeorhetic alterations  
1472 observed during the gestation of zebu beef heifers to benefit fetal development.

1473

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1481

## 1482 **References**

1483 Aschenbach, J.R., Penner, G.B., Stumpff, F., Gabel, G., 2011. Ruminant Nutrition  
1484 Symposium: Role of fermentation acid absorption in the regulation of ruminal  
1485 pH. *J. Anim. Sci.* 89, 1092-1107.

1486 Bach, A., Guasch, I., Elcoso, G., Chaucheyras-Durand, F., Castex, M., Fabregas, F.,  
1487 Garcia-Fruitos, E., Aris, A., 2018. Changes in gene expression in the rumen and  
1488 colon epithelia during the dry period through lactation of dairy cows and effects  
1489 of live yeast supplementation. *J. Dairy Sci.* 101, 2631-2640.

1490 Baldwin, R.J., 1999. The proliferative actions of insulin, insulin-like growth factor-I,  
1491 epidermal growth factor, butyrate and propionate on ruminal epithelial cells in  
1492 vitro. *Small Ruminant Res.* 32, 261-268.

1493 Baldwin, R.L., Wu, S., Li, W., Li, C., Bequette, B.J., Li, R.W., 2012. Quantification of  
1494 transcriptome responses of the rumen epithelium to butyrate infusion using  
1495 RNA-seq technology. *Gene Regul. Syst. Bio.* 6, 67-80.

1496 Bauman, D.E., Currie, W.B., 1980. Partitioning of nutrients during pregnancy and  
1497 lactation: a review of mechanisms involving homeostasis and homeorhesis. *J.*  
1498 *Dairy Sci.* 63, 1514-1529.

1499 Bell, A.W., 1995. Regulation of organic nutrient metabolism during transition from late  
1500 pregnancy to early lactation. *J. Anim. Sci.* 73, 2804-2819.

1501 Brosh, A., Aharoni, Y., Holzer, Z., 2002. Energy expenditure estimation from heart rate:  
1502 validation by long-term energy balance measurement in cows. *Livest. Prod. Sci.*  
1503 77, 287-299.

- 1504 Connor, E.E., Li, R.W., Baldwin, R.L., Li, C., 2010. Gene expression in the digestive  
1505 tissues of ruminants and their relationships with feeding and digestive  
1506 processes. *Animal* 4, 993-1007.
- 1507 Die, J.V., Baldwin, R.L., Rowland, L.J., Li, R., Oh, S., Li, C., Connor, E.E., Ranilla, M.-  
1508 J.J., 2017. Selection of internal reference genes for normalization of reverse  
1509 transcription quantitative polymerase chain reaction (RT-qPCR) analysis in the  
1510 rumen epithelium. *PLoS One* 12, e0172674.
- 1511 Forbes, J.M., 1968. The physical relationships of the abdominal organs in the pregnant  
1512 ewe. *J. Agr. Sci.* 70, 171-177.
- 1513 Freetly, H., Nienaber, J., Brown-Brandl, T., 2008. Partitioning of energy in pregnant  
1514 beef cows during nutritionally induced body weight fluctuation. *J. Anim. Sci.* 86,  
1515 370-377.
- 1516 Ghizzi, L.G., Del Valle, T.A., Takiya, C.S., da Silva, G.G., Zilio, E.M., Grigoletto, N.T.,  
1517 Martello, L.S., Rennó, F.P., 2018. Effects of functional oils on ruminal  
1518 fermentation, rectal temperature, and performance of dairy cows under high  
1519 temperature humidity index environment. *Anim. Feed Sci. Tech.* 246, 158-166.
- 1520 Gionbelli, M.P., Valadares Filho, S.C., Duarte, M.S. 2016. Nutritional requirements for  
1521 pregnant and non-pregnant beef cows, in: Valadares Filho, S.C. et al. (Eds.),  
1522 *Nutrient Requirements of Zebu and Crossbred Cattle - BR-CORTE 3.0.*  
1523 *Suprema, Viçosa, Brazil*, pp. 251-272.
- 1524 Hanks, D.R., Judkins, M.B., McCracken, B.A., Holcombe, D.W., Krysl, L.J., Park, K.K.,  
1525 1993. Effects of pregnancy on digesta kinetics and ruminal fermentation in beef  
1526 cows. *J. Anim. Sci.* 71, 2809-2814.
- 1527 Hansen, P.J., 2004. Physiological and cellular adaptations of zebu cattle to thermal  
1528 stress. *Anim. Reprod. Sci.* 82, 349-360.



- 1529 Hare, K.S., Wood, K.M., Acton, K., Fitzsimmons, C., Penner, G.B., 2019.  
1530 Oversupplying metabolizable protein in late gestation for beef cattle: effects on  
1531 prepartum BW, ruminal fermentation, nitrogen balance, and skeletal muscle  
1532 catabolism. *J. Anim. Sci.* 97, 407-423.
- 1533 Kantyka, T., Fischer, J., Wu, Z., Declercq, W., Reiss, K., Schroder, J.M., Meyer-Hoffert,  
1534 U., 2011. Inhibition of kallikrein-related peptidases by the serine protease  
1535 inhibitor of Kazal-type 6. *Peptides* 32, 1187-1192.
- 1536 Kern, R.J., Lindholm-Perry, A.K., Freetly, H.C., Kuehn, L.A., Rule, D.C., Ludden, P.A.,  
1537 2016a. Rumen papillae morphology of beef steers relative to gain and feed  
1538 intake and the association of volatile fatty acids with kallikrein gene expression.  
1539 *Livest. Sci.* 187, 24-30.
- 1540 Kern, R.J., Lindholm-Perry, A.K., Freetly, H.C., Snelling, W.M., Kern, J.W., Keele,  
1541 J.W., Miles, J.R., Foote, A.P., Oliver, W.T., Kuehn, L.A., Ludden, P.A., 2016b.  
1542 Transcriptome differences in the rumen of beef steers with variation in feed  
1543 intake and gain. *Gene* 586, 12-26.
- 1544 Kvidera, S., Horst, E., Abuajamieh, M., Mayorga, E., Fernandez, M.S., Baumgard,  
1545 L.H., 2017. Glucose requirements of an activated immune system in lactating  
1546 Holstein cows. *J. Dairy Sci.* 100, 2360-2374.
- 1547 Laarman, A., Pederzolli, R.-L., Wood, K., Penner, G., McBride, B.W., 2016. Effects of  
1548 subacute ruminal acidosis and low feed intake on short-chain fatty acid  
1549 transporters and flux pathways in Holstein steers. *J. Anim. Sci.* 94, 3729-3737.
- 1550 Laarman, A.H., Dionissopoulos, L., AlZahal, O., Greenwood, S.L., Steele, M.A.,  
1551 McBride, B.W., 2013. Butyrate and subacute ruminal acidosis affect abundance  
1552 of membrane proteins involved with proton and short chain fatty acid transport  
1553 in the rumen epithelium of dairy cows. *Am. J. Anim. Vet. Sci.* 8, 220-229.

- 1554 Lima, S., Stafuzza, N.B., Pires, B.V., Bonilha, S.F.M., Cyrillo, J., Negrao, J.A., Paz,  
1555 C.C.P., 2020. Effect of high temperature on physiological parameters of Nelore  
1556 (Bos taurus indicus) and Caracu (Bos taurus taurus) cattle breeds. Trop. Anim.  
1557 Health Prod. 52, 2233-2241.
- 1558 Lopes, R.C., Sampaio, C.B., Trece, A.S., Teixeira, P.D., Gionbelli, T.R.S., Santos,  
1559 L.R., Costa, T.C., Duarte, M.S., Gionbelli, M.P., 2020. Impacts of protein  
1560 supplementation during late gestation of beef cows on maternal skeletal muscle  
1561 and liver tissues metabolism. Animal 14(9), 1867-1875
- 1562 Melo Costa, C.C., Maia, A.S.C., Nascimento, S.T., Nascimento, C.C.N., Neto, M.C.,  
1563 de Franca Carvalho Fonseca, V., 2018. Thermal balance of Nelore cattle. Int.  
1564 J. Biometeorol. 62, 723-731.
- 1565 Meyer, A.M., Reed, J.J., Vonnahme, K.A., Soto-Navarro, S.A., Reynolds, L.P., Ford,  
1566 S.P., Hess, B.W., Caton, J.S., 2010. Effects of stage of gestation and nutrient  
1567 restriction during early to mid-gestation on maternal and fetal visceral organ  
1568 mass and indices of jejunal growth and vascularity in beef cows. J. Anim. Sci.  
1569 88, 2410-2424.
- 1570 Milan, H., Maia, A., Gebremedhin, K.G., 2016. Device for measuring respiration rate of  
1571 cattle under field conditions. J. Anim. Sci. 94, 5434-5438.
- 1572 Minuti, A., Palladino, A., Khan, M., Alqarni, S., Agrawal, A., Piccioli-Capelli, F., Hidalgo,  
1573 F., Cardoso, F., Trevisi, E., Looor, J.J., 2015. Abundance of ruminal bacteria,  
1574 epithelial gene expression, and systemic biomarkers of metabolism and  
1575 inflammation are altered during the peripartal period in dairy cows. J. Dairy Sci.  
1576 98, 8940-8951.
- 1577 Pfaffl, M.W., 2001. A new mathematical model for relative quantification in real-time  
1578 RT-PCR. Nucleic Acids Res. 29, e45-e45.

- 1579 Reynolds, J., Foote, A., Freetly, H., Oliver, W., Lindholm-Perry, A.K., 2017.  
1580 Relationships between inflammation-and immunity-related transcript  
1581 abundance in the rumen and jejunum of beef steers with divergent average daily  
1582 gain. *Anim. Genet.* 48, 447-449.
- 1583 Reynolds, L., Ferrell, C., Robertson, D.A., Ford, S.P., 1986. Metabolism of the gravid  
1584 uterus, foetus and utero-placenta at several stages of gestation in cows. *J. Agr.*  
1585 *Sci.* 106, 437-444.
- 1586 Ribeiro, R.C.O., Villela, S.D.J., Valadares Filho, S.C, Santos, S.A., Ribeiro, K.G.,  
1587 Detmann, E., Zanetti, D., Martins, P.G.M.A., 2015. Effects of roughage sources  
1588 produced in a tropical environment on forage intake, and ruminal and microbial  
1589 parameters. *J. Anim. Sci.* 93, 2363-2374.
- 1590 Rotta, P.P., Valadares Filho, S.C., Gionbelli, T.R.S., Costa e Silva, L.F., Engle, T.E.,  
1591 Marcondes, M.I., Campos, M.M., Menezes, A.C.B., Lobo, A.A.G., 2015. Effects  
1592 of day of gestation and feeding regimen in Holsteinx Gyr cows: II. Maternal and  
1593 fetal visceral organ mass. *J. Dairy Sci.* 98, 3211-3223.
- 1594 Scheaffer, A., Caton, J., Bauer, M., Redmer, D., Reynolds, L.P., 2003. The effect of  
1595 pregnancy on visceral growth and energy use in beef heifers. *J. Anim. Sci.* 81,  
1596 1853-1861.
- 1597 Scheaffer, A., Caton, J., Bauer, M., Reynolds, L., 2001. Influence of pregnancy on body  
1598 weight, ruminal characteristics, and visceral organ mass in beef heifers. *J. Anim.*  
1599 *Sci.* 79, 2481-2490.
- 1600 Stanley, T., Cochran, R., Vanzant, E., Harmon, D., Corah, L., 1993. Periparturient  
1601 changes in intake, ruminal capacity, and digestive characteristics in beef cows  
1602 consuming alfalfa hay. *J. Anim. Sci.* 71, 788-795.

- 1603 Steele, M., Schiestel, C., AlZahal, O., Dionissopoulos, L., Laarman, A., Matthews, J.,  
1604 McBride, B.W., 2015. The periparturient period is associated with structural and  
1605 transcriptomic adaptations of rumen papillae in dairy cattle. *J. Dairy Sci.* 98,  
1606 2583-2595.
- 1607 Trenk, L., Kuhl, J., Aurich, J., Aurich, C., Nagel, C., 2015. Heart rate and heart rate  
1608 variability in pregnant dairy cows and their fetuses determined by fetomaternal  
1609 electrocardiography. *Theriogenology* 84, 1405-1410.
- 1610 Valadares Filho, S.C., Costa e Silva, L.F., Gionbelli, M.P., Rotta, P.P., Marcondes,  
1611 M.I., Chizzotti, M.L., Prados, L.F. 2016. *Nutrient Requirements of Zebu and*  
1612 *Crossbred Cattle - BR-CORTE 3.0*, third ed. Suprema, Viçosa, Brazil.
- 1613 Veerkamp, R.F., Coffey, M.P., Berry, D.P., De Haas, Y., Strandberg, E., Bovenhuis,  
1614 H., Calus, M., Wall, E., 2012. Genome-wide associations for feed utilisation  
1615 complex in primiparous Holstein–Friesian dairy cows from experimental  
1616 research herds in four European countries. *Animal* 6, 1738-1749.
- 1617 Wood, K., Awda, B., Fitzsimmons, C., Miller, S., McBride, B., Swanson, K., 2013.  
1618 Influence of pregnancy in mid-to-late gestation on circulating metabolites,  
1619 visceral organ mass, and abundance of proteins relating to energy metabolism  
1620 in mature beef cows. *J. Anim. Sci.* 91, 5775-5784.
- 1621 Zhang, S., Albornoz, R., Aschenbach, J., Barreda, D., Penner, G.B., 2013. Short-term  
1622 feed restriction impairs the absorptive function of the reticulo-rumen and total  
1623 tract barrier function in beef cattle. *J. Anim. Sci.* 91, 1685-1695.

**Table 1**

Sequences (5' to 3') and efficiencies of the primers used in quantitative real-time PCR.

Gene	Symbol	Forward (F) and reverse (R)	Access number	Amplicon size (bp)	R <sup>2</sup>	Efficiency
<i>PCNA</i>	PCNA	F: GCTACACTTTCCTCAGTCCTTC R: GCCTCCAGCACTTTCTTCA	XM_019973361.1	105	0.97	1.00
<i>CASP3</i>	CASP3	F: GAGACGGGTTGAGGACAATAAG R: TGACAGAAGAGCCCTTTAGATATTC	XM_019953295.1	96	0.99	1.03
<i>KLK10</i>	KLK10	F: GGGTGGTGA ACTCTGACTAAAT R: GCAAAGGGTGGTTAGGATTAGA	XM_019978443.1	100	0.99	1.03
<i>KLK9</i>	KLK9	F: TCTCTGAGTCACCAGGA ACT R: GGGAAGCACCTGAAGCTATT	XM_019978450.1	97	0.93	1.04
<i>SLC16A1</i>	MCT1	F: TGTGGGACTGAAGGGTAAATG R: CCTGGTATGATTCCCACAGAAA	XM_019956796.1	114	0.98	1.00
<i>SLC16A3</i>	MCT4	F: CTGGTGCTGGGTA ACTTCTT R: GTTCTTCTCAGGCTCTGTCTTC	XM_019980902.1	138	0.98	0.93
<i>SLC26A3</i>	DRA	F: GGATTTCTCTTGGAGCCTCTAC R: CTTTCGCCACAATCTTCGTATTT	XM_019958684.1	108	0.99	0.97
<i>SLC26A6</i>	PAT1	F: GGGACTGAGCTAGAGGATACA R: CAGGATGAGGGTGTGGAAAT	NM_001076852.2	114	0.97	1.03
<i>SLC9A1</i>	NHE1	F: CCCATTCTATTCCCTCCTCTGTC R: AGAGGGACCAGGACCTATTT	XM_019980388.1	124	0.99	0.99
<i>SLC5A1</i>	SGLT1	F: TCCTGACTGGGTTTGCTTTC R: TGACGGTGGTGGTTCCATAAG	XM_019977410.1	105	0.98	0.98

*PCNA* = Proliferating cell nuclear antigen; *CASP3* = Caspase 3; *KLK10* = Kallikrein-related peptidase 10; *KLK9* = Kallikrein-related peptidase 9; *SLC16A1* (MCT1) = Monocarboxylate transporter 1; *SLC16A3* (MCT4) = Monocarboxylate transporter 4; *SLC26A3*

(DRA) = Down-regulated in adenoma; *SLC26A6* (PAT1) = Putative anion transporter 1; *SLC9A1* (NHE1) = Sodium-hydrogen antiporter 1; *SLC5A1* (SGLT1) = Sodium/glucose cotransporter 1.

**Table 2**

Effect of physiological status (PS) and days of pregnancy (DOP) on respiratory (RR; breaths/min) and heart (HR; beats/min) rates of beef heifers.

Item	Physiological status		SEM	P-value		
	Non-pregnant	Pregnant		PS	DOP	PSxDOP
Before morning feeding						
RR	23.2	24.6	1.1	0.35	0.44	0.79
HR	70.2	74.2	1.1	0.01	0.04	0.08
Four hours after morning feeding						
RR	30.6	32.1	1.6	0.47	0.01	0.96
HR	74.8	82.0	1.8	<0.01	<0.01	0.63

PSxDOP = interaction between physiological status and days of pregnancy.

**Table 3**

Effect of physiological status (PS) and days of pregnancy (DOP) on blood metabolites levels of beef heifers.

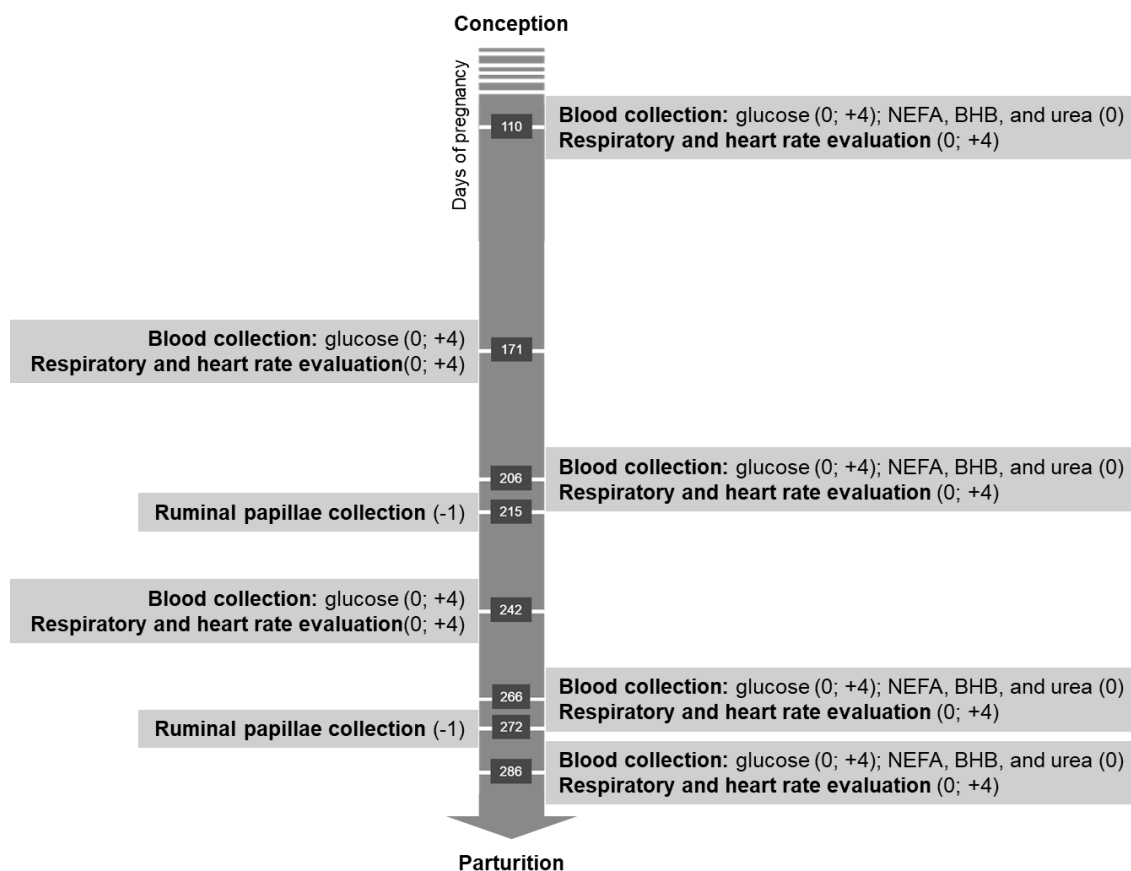
Item	Physiological status		SEM	P-value		
	Non-pregnant	Pregnant		PS	DOP	PSxDOP
Before morning feeding						
NEFA, $\mu\text{mol/L}$	202	211	54	0.82	<0.01	0.11
BHB, $\mu\text{mol/L}$	348	364	42	0.77	0.24	0.67
Urea, mg/dL	21.1	21.5	2.1	0.78	<0.01	0.03
Glucose (mg/dL	100	96.6	3.7	0.41	0.03	0.05
Four hours after morning feeding						
Glucose, mg/dL	97.2	96.5	2.7	0.82	0.20	0.17

NEFA = non-esterified fatty acids; BHB = beta-hydroxybutyrate; PSxDOP = interaction

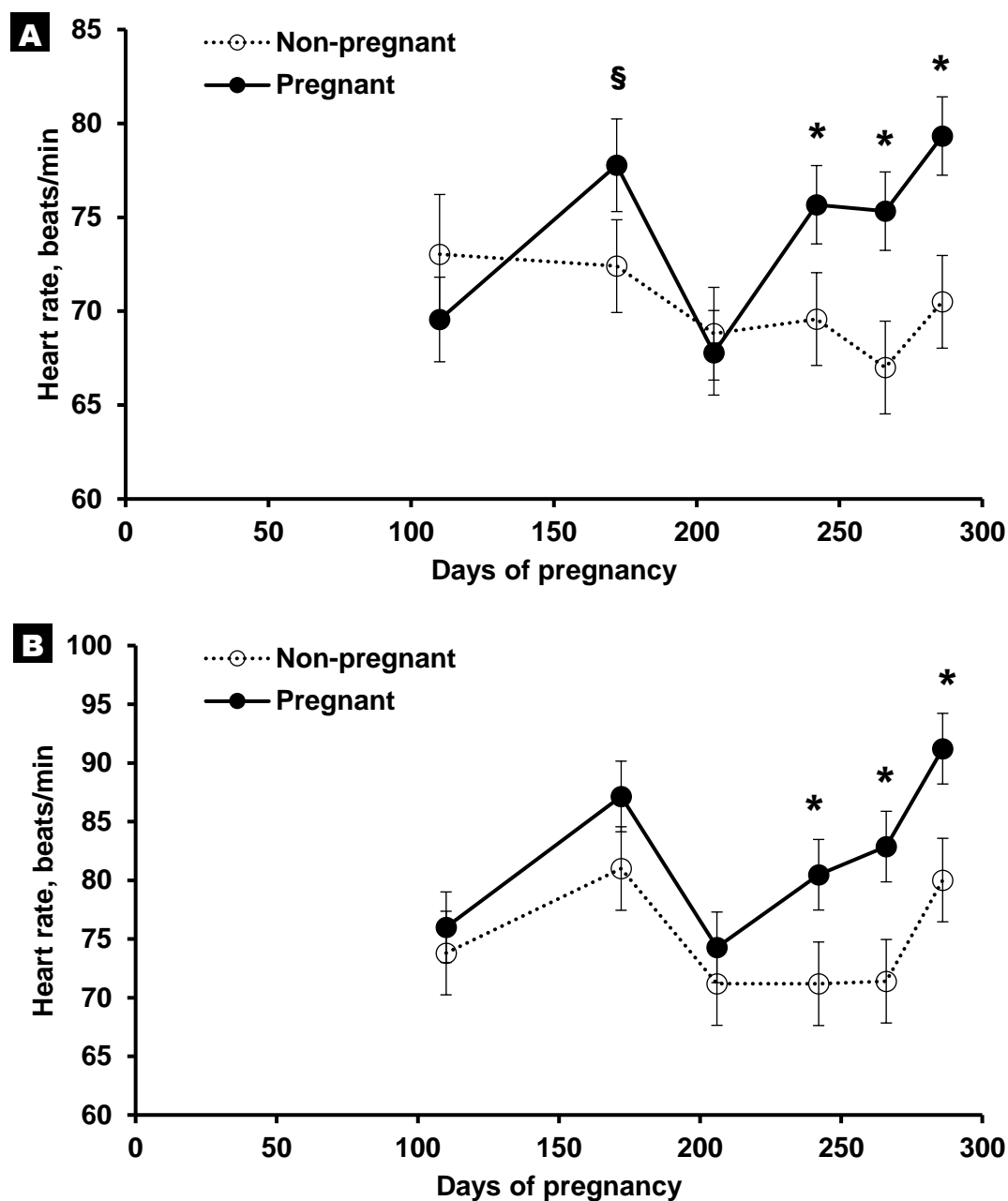
between physiological status and days of pregnancy.



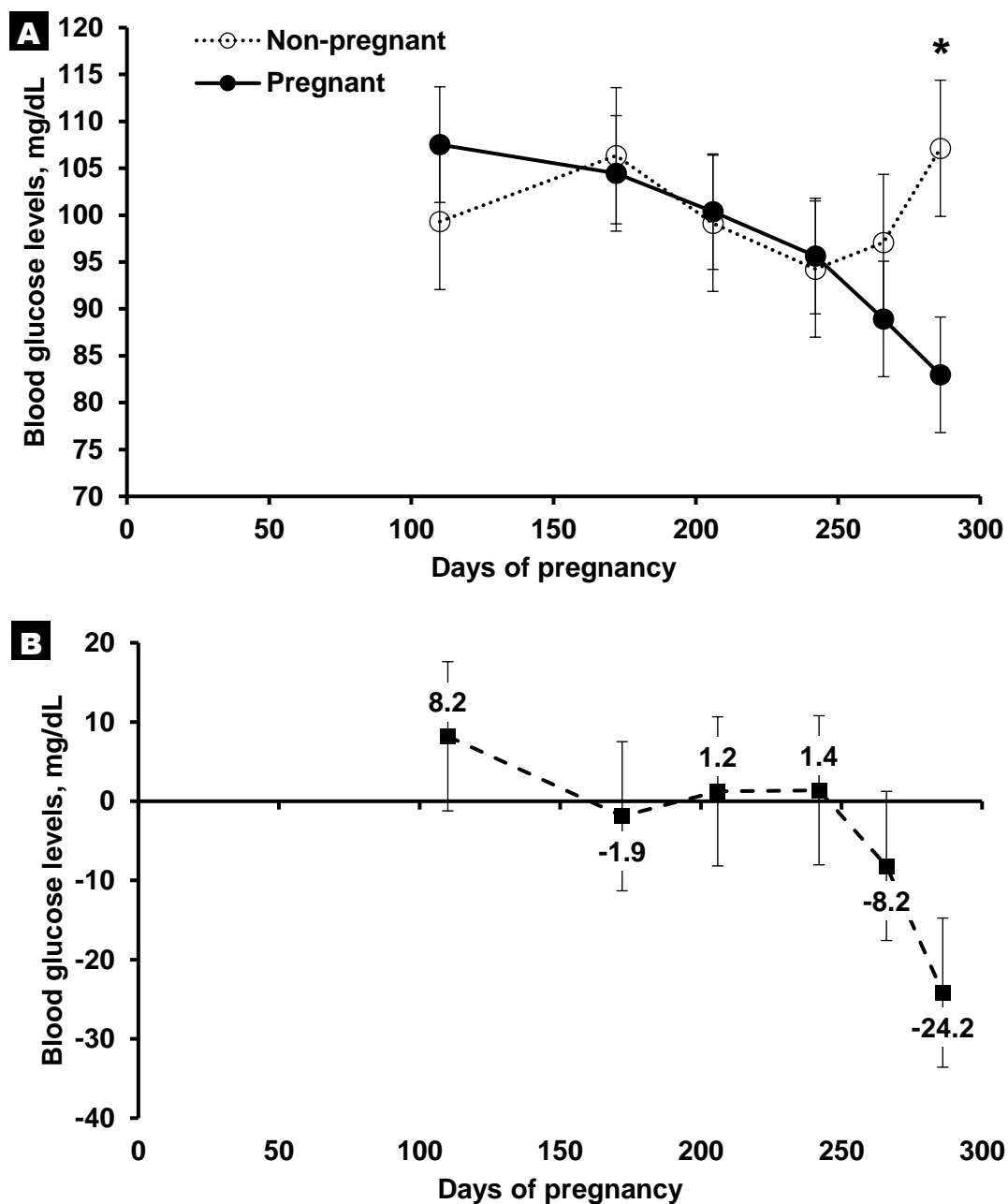
## Figures



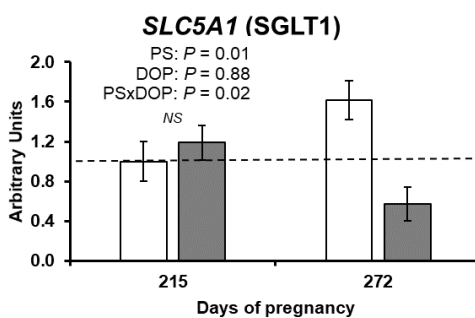
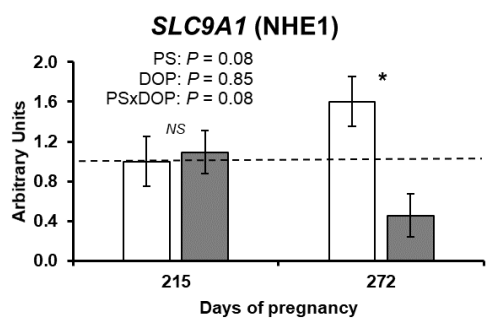
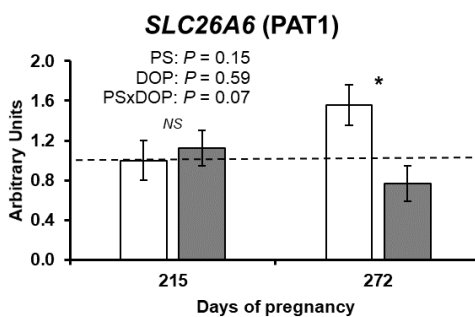
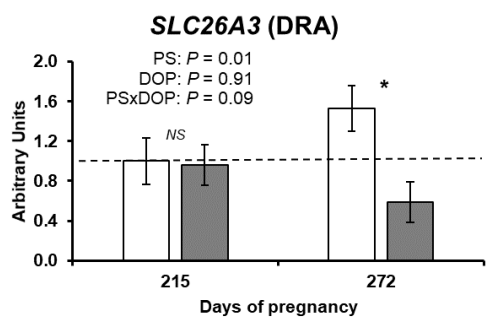
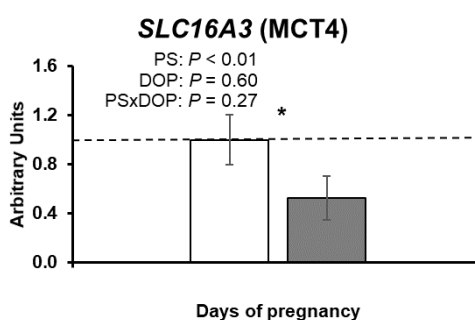
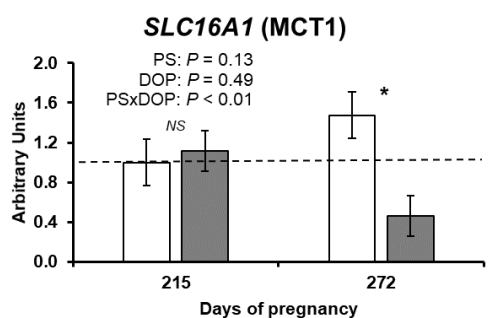
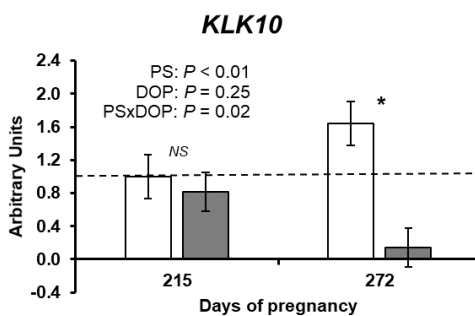
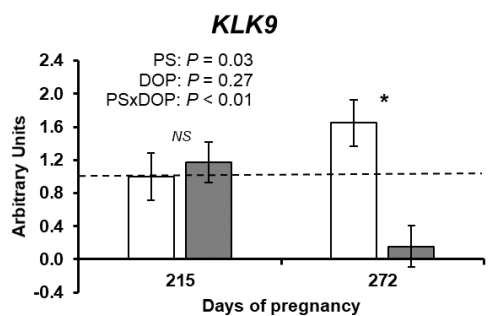
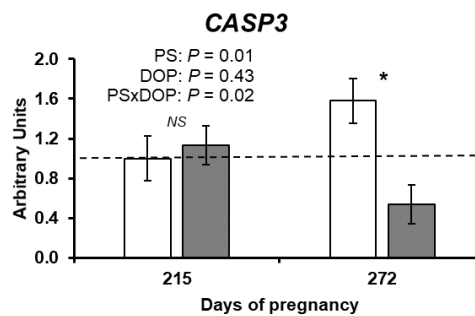
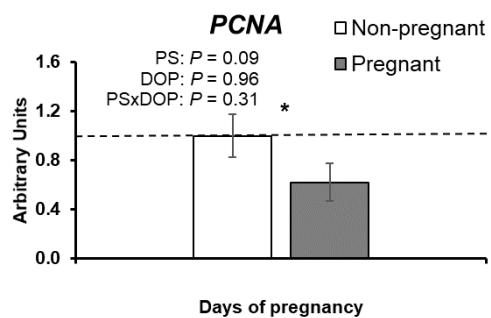
**Fig. 1.** Timeline of data collections during pregnancy of beef heifers. Collection times: where -1: one hour before the morning feeding, 0: immediately before the morning feeding, and +4: four hours after the morning feeding. NEFA = non-esterified fatty acids; BHB = beta-hydroxybutyrate.



**Fig. 2** Effect of physiological status and days of pregnancy on the heart rate of beef heifers. **(A)** Heart rates of pregnant and non-pregnant beef heifers (beats/min) over days assessed just before morning feeding. **(B)** Heart rates of pregnant and non-pregnant beef heifers (beats/min) over days assessed four hours after the morning feeding. §  $P < 0.15$ ; \*  $P \leq 0.10$ .



**Fig. 3** Effect of physiological status and days of pregnancy on blood glucose concentration of beef heifers. **(A)** Blood glucose concentration in pregnant and non-pregnant beef heifers (mg/dL) over days. **(B)** Estimated blood glucose concentration (mg/dL) between pregnant and non-pregnant beef heifers over days. \*  $P \leq 0.10$ .



**Fig. 4** Relative gene expression in ruminal papillae of beef heifers according to physiological status (PS), days of pregnancy (DOP) and their interaction (PS×DOP). Proliferating cell nuclear antigen (*PCNA*), caspase 3 (*CASP3*), kallikrein-related peptidase 9 (*KLK9*), kallikrein-related peptidase 10 (*KLK10*), monocarboxylate transporter 1 (*SLC16A1/MCT1*), monocarboxylate transporter 4 (*SLC16A3/MCT4*), down-regulated in adenoma (*SLC26A3/DRA*), putative anion transporter 1 (*SLC26A6/PAT1*), sodium-hydrogen antiporter 1 (*SLC9A1/NHE1*), sodium/glucose cotransporter 1 (*SLC5A1/SGLT1*), NS = Non-significant. \*  $P < 0.10$ .